

# Pharmacognostical Screening and Evaluation of In Vitro Anti-Glycation Activity of Eucalyptus globulus Leaf Extract

Dipak Bankar, Mayur Barkade, Dr. Nitin Mali

Vidya Niketan College of Pharmacy, Lakhewadi, Indapur

dipak2003bankar@gmail.com, mayurbarkade2021@gmail.com, nitinmalivncop@gmail.com

**Abstract:** *The non-enzymatic interaction between reducing sugars and proteins, lipids, or nucleic acids produces advanced glycation end products (AGEs), which accelerate the development of chronic conditions including diabetes, heart disease, and aging-related problems. In this work, the antiglycation efficacy of Eucalyptus globulus leaf extract was assessed, and the phytochemical components responsible for its therapeutic actions were examined. Eucalyptus globulus leaves were gathered, dried, pulverized, and extracted using the Soxhlet extraction technique with methanol and ethanol as solvents. While determining physicochemical characteristics including ash value and drying loss, preliminary phytochemical screening was carried out to find beneficial chemicals. An in vitro titration approach utilizing fructose-protein reaction mixtures at various extract concentrations (20–100 µg/mL) was used to assess antiglycation activity.*

*Important secondary metabolites, such as alkaloids, flavonoids, tannins, saponins, terpenoids, and phenolic chemicals, were found during phytochemical screening, suggesting significant therapeutic potential. The crude drug's acceptable quality and moisture content were confirmed by the ash value and drying loss of 5% and 10%, respectively. The antiglycation test showed concentration-dependent inhibition, with glycation reactions being considerably decreased as extract concentration increased. At a dose of 100 µg/mL, the maximum antiglycation activity of 50% inhibition was noted. Because of their antioxidant qualities, phytoconstituents—especially flavonoids and phenolic compounds—may have a useful role in the decrease of titration volume and glycation end-product production. According to the study's findings, Eucalyptus globulus has a good antiglycation potential and might be used as a natural medicinal source to prevent glycation-related problems connected with aging and diabetes...*

**Keywords:** *advanced glycation end products*

## I. INTRODUCTION

Reducing sugars combine with proteins, lipids, or nucleic acids in a non-enzymatic biochemical process called advanced glycation, which produces toxic substances called advanced glycation end products (AGEs). Numerous chronic illnesses, including diabetes mellitus, cardiovascular problems, neurodegenerative diseases, renal dysfunction, and aging-related issues, are closely linked to the formation of AGEs. AGEs interfere with normal physiological processes by causing oxidative stress, inflammation, and cellular damage. As a result, glycation inhibition or prevention has drawn a lot of interest in biological and pharmacological research.

Because of their rich phytochemical composition, safety, and affordability, natural medicinal plants have been extensively investigated as alternative sources of therapeutic chemicals. Flavonoids, tannins, alkaloids, terpenoids, and phenolic compounds are examples of plant-derived bioactive substances with antioxidant, anti-inflammatory, antibacterial, and antiglycation qualities. These phytochemicals play a critical role in reducing the production of AGEs by inhibiting oxidative damage and neutralizing free radicals.



*Eucalyptus globulus*, sometimes referred to as Tasmanian blue gum or blue gum, is a member of the Myrtaceae family and is well known for its industrial and medicinal use. The plant is widely grown across many tropical and subtropical parts of the world, but it is native to Australia and Tasmania. The therapeutic benefit of eucalyptus leaves is attributed to their abundance of essential oils, particularly eucalyptol (1,8-cineole), as well as a variety of phytoconstituents, including flavonoids, tannins, terpenoids, alkaloids, and phenolic compounds. The herb has long been used to treat fever, wounds, inflammation, microbial infections, and respiratory conditions.

## II. MATERIAL AND METHODS:

Plant Profile: *Eucalyptus globulus*



Fig.no.1 *Eucalyptus globulus* leaves

Common Name: Blue Gum, Tasmanian Blue Gum

Botanical Name: *Eucalyptus globulus*

Family: Myrtaceae

Description: The tall evergreen *Eucalyptus globulus*, sometimes called Blue Gum, is a member of the Myrtaceae family of trees. Originally from Australia and Tasmania, it has extensive industrial and therapeutic value, making it a popular choice for cultivation in temperate, tropical, and subtropical climates. The tree has a straight trunk covered with smooth, bluish-grey bark that peels off in strips and can reach heights of 30 to 70 meters.

The presence of essential oils gives the leaves their fragrant quality. Mature leaves are long, thin, lance-shaped, and dark green, but juvenile leaves are opposite, wide, and bluish-green. The creamy white, nectar-rich blooms are solitary. Often referred to as a "gumnut," the fruit is a woody capsule that contains many tiny seeds.

The plant is frequently planted beside highways, on plantations, and in forested regions since it grows well in sunny, well-drained soils. It is prized for its lumber, essential oil, and therapeutic qualities

Chemical Constituents:

- Essential oils (mainly eucalyptol/1,8-cineole) – major active constituent responsible for medicinal properties
- Flavonoids – antioxidant compounds
- Tannins – possess antimicrobial and astringent properties
- Phenolic compounds – contribute to antioxidant activity



- Terpenoids and monoterpenes – responsible for anti-inflammatory effects
- Alkaloids – present in small quantities
- Resins and waxes – present in leaves and bark
- Organic acids and carbohydrates – primary metabolites

**Uses:**

**1. Medicinal Uses**

- Antiglycation activity: aids in preventing the production of AGEs, or advanced glycation end products.
- Antioxidant: flavonoids and phenolic substances scavenge free radicals.
- Anti-inflammatory: lessens inflammation in tests
- Antimicrobial: efficient against certain fungi and bacteria
- Expectorant and decongestant: often used for respiratory conditions, colds, and coughs
- Eucalyptus oil is an antiseptic used in preparations for disinfectants and wound cleansing.

**2. Applications in Industry**

- Production of essential oils: Eucalyptus oil is mostly derived from leaves.
- The timber and paper business uses wood for building, pulp, and paper.
- Fuelwood: Due to its quick growth, it is frequently utilized as firewood.
- Products that repel insects, such as oil used in sprays and repellents for mosquitoes

**3. Customary and Folk Applications**

- Inhaling steam from leaves to relieve congestion in the nose and cough
- External use of leaf paste and oil for mild injuries and sore muscles
- Used in conventional treatments for respiratory infections and fever

**4. Decorative Use**

- Because of its towering stature and fragrant leaf, it is grown as a shade and decorative tree.

**III. METHODOLOGY**

**EXTRACTION OF PLANT EXTRACT:**

**Step 1: Weighing**

Accurately weigh 50 g of powdered leaf sample using an electronic weighing balance.

**Step 2: Filling the Thimble**

Transfer the powder into a cellulose extraction thimble.

The thimble prevents solid particles from mixing with solvent.

**Step 3: Assembly of Soxhlet Apparatus**

The Soxhlet apparatus consists of:

Round-bottom flask

Extraction chamber

Condenser

Heating mantle

**Step 4: Addition of Solvent**

Add 300–500 mL methanol or ethanol into the round-bottom flask.

Why methanol/ethanol?

These solvents effectively extract:

Phenols

Flavonoids

Alkaloids



Glycosides

Step 5: Heating

Heat the solvent at 60–70°C.

The solvent evaporates and moves upward into the condenser.

Step 6: Condensation

Cold water circulating through the condenser cools the solvent vapor.

The vapor condenses into liquid form and drips onto the plant powder.

Step 7: Extraction Cycle

The chamber fills with solvent.

When the solvent reaches siphon level, it automatically drains back into the flask carrying dissolved phytochemicals.

This cycle repeats continuously for 6–8 hours.

Usually 8–12 cycles are sufficient.

Step 8: Completion

Extraction is complete when the solvent in siphon tube becomes colorless.

Step 9: Filtration

The extract is filtered using Whatman filter paper No.1.

Purpose: To remove suspended particles.

Step 10: Concentration of Extract

The filtrate contains solvent + extracted phytochemicals.

It is concentrated by:

Water Bath Method

Heat at 40–50°C until solvent evaporates.

OR

Rotary Evaporator

Used under reduced pressure for efficient solvent removal.

The concentrated crude extract is:

Sticky/semi-solid

Dark green/brown

The extract is weighed and percentage yield is calculated:

Step 11: Storage of Extract

The crude extract is transferred into sterile airtight containers.

Stored at 4°C in refrigerator.

Purpose: Prevents:

Oxidation

Microbial growth

Chemical degradation





Fig.no. 2 Soxhlet Apparatus

**Preliminary Phytochemical Screening:**

**A. Alkaloid test:**

TEST	OBSERVATION
Mayer's Test Take 2-3 ml of plant extract in a test tube. Add few drops of dil.HCL and warm gently. Add Mayer's reagent	Pale yellow precipitate
Dragendorff's Test Take 2-3 ml of plant extract in a test tube. Add few drops of dil.HCL and warm gently. Add Dragendorff's reagent	Reddish-brown precipitate
Wagner's Test Take 2-3 ml of plant extract in a test tube. Add few drops of dil.HCL and warm gently. Add Wagner's reagent	Reddish-brown precipitate

Table No.1 Alkaloid test

**B. Flavonoids Test:**

Shinoda Test Take 2-3 ml of plant extract in a test tube.	Pink colour are observed
---	--------------------------



Add small Zinc Dust	
Add few drops of dil.HCL	

Table No.2 Flavonoids Test

**C. Saponin Test:**

Foam Test Take 1ml plant extract. Add 5-10 ml distilled water. Shake vigorously for 30 sec. Let it stand for 10-15 min.	Persistent foam (>1cmheight)
---	---------------------------------

Table No.3 Saponin Test

**D. Tannins Test:**

Take 2ml plant extract. Add 2-3 drops of FeCl <sub>3</sub> solution.	Black color observed
---	----------------------

Table No.4 Tannin Test

**E. Terpenoids Test**

Salkowski Test Take 2ml plant extract. Add 2ml chloroform. Carefully add 2-3 ml conc. H <sub>2</sub> SO <sub>4</sub> along the side of the test tube.	Brown ring at junction
---	------------------------

Table No.5 Terpenoids Test:

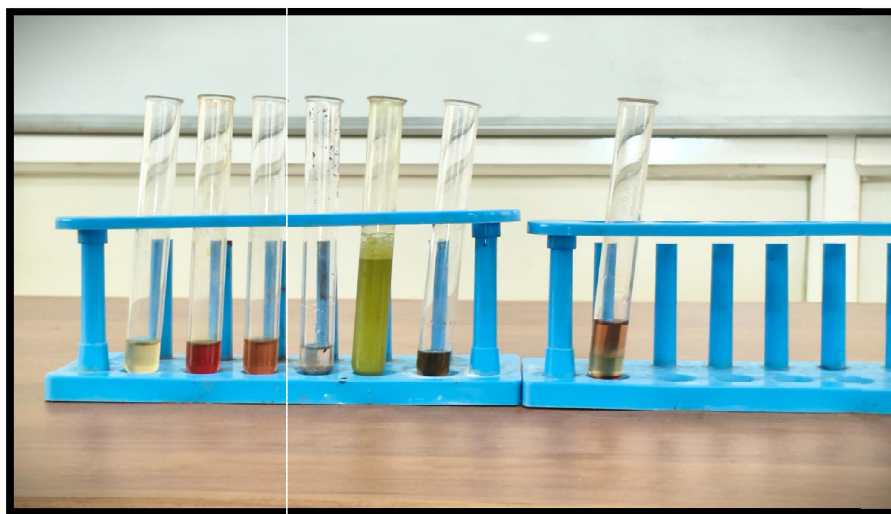


Fig.no.3 Preliminary Phytochemical Screening

**ASH VALUE:**

The ash value of Eucalyptus globulus is determined by the incineration (heating) method used in pharmacognosy to measure the inorganic residue left after burning the plant material.



Principle:

When the powdered drug is heated at high temperature, all organic matter burns off and the remaining residue is called ash. It indicates the presence of:

- Natural mineral content
- Adhering dirt, sand, or impurities
- Adulteration
- Heating / Incineration Method for Ash Value Determination

Procedure:

1. Weigh about 2–3 g of air-dried powdered Eucalyptus globulus.
2. Place it in a previously ignited and weighed silica crucible.
3. Heat gently at first to avoid loss due to smoke.
4. Then incinerate in a muffle furnace at about 450–600°C until the sample becomes white, indicating absence of carbon.
5. Cool in a desiccator and weigh the ash.

Formula:

$$\text{Ash Value (\%w/w)} = \frac{\text{Weight of Ash}}{\text{Weight of air dried crude drug}} \times 100$$

### LOSS OF DRYING

Principle:

The powdered drug is heated at 105°C until constant weight is obtained. The weight loss indicates moisture and volatile components removed during drying.

Procedure:

1. Weigh about 2–5 g of powdered drug.
2. Place in a previously dried and weighed evaporating dish.
3. Dry in hot air oven at 105°C.
4. Cool in desiccator and weigh.
5. Repeat until constant weight is obtained.

Formula:

$$\text{Loss of Drying (\%w / w)} = \frac{\text{Initial weight} - \text{Final weight}}{\text{Initial weight}} \times 100$$

### PHARMACOLOGICAL ACTIVITY

Anti-Glycation Assay By Titration Method

Principle-

Glycation reaction leaves free reducing sugars in solution. These are estimated by titration. The lower the titration volume, the higher the anti-glycation activity.

Key Idea-Instead of control flask:

Use 0 µg/mL (blank mixture) as reference titre value

Then compare all test samples against it.

Materials-

Fructose (1% or 0.5%), Gelatin, Plant extract (20–100 µg/mL), Fehling's solution A & B, Methylene blue indicator, Distilled water

Procedure-

1. Prepare Reaction Mixture

For each tube:

2 mL fructose solution



2 mL Gelatin solution  
1 mL extract (or water for blank)

2. Set of Tubes

You prepare:

TUBE	EXTRACT
Blank( $V_0$ )	0 $\mu\text{g}/\text{Ml}$
T1	20 $\mu\text{g}/\text{mL}$
T2	40 $\mu\text{g}/\text{mL}$
T3	60 $\mu\text{g}/\text{mL}$
T4	80 $\mu\text{g}/\text{mL}$
T5	100 $\mu\text{g}/\text{mL}$

Incubation-

60°C water bath for 1 hour.

Titration-

For each tube:

1. Take Fehling's A + B (5 mL each)
2. Boil gently
3. Titrate with incubated sample
4. Add methylene blue near endpoint
5. Record volume

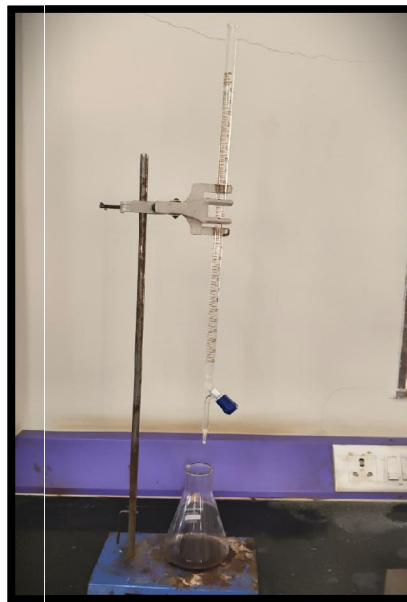


Fig.no.4 Anti-Glycation Assay By Titration Method



**IV. RESULTS AND DISCUSSION:**

**PRELIMINARY TESTS:**

Alkaloid test:

TEST	OBSERVATION	INFERENCE
Mayer's Test Take 2-3 ml of plant extract in a test tube. Add few drops of dil.HCL and warm gently. Add Mayer's reagent	Pale yellow precipitate	Alkaloid is present.
Dragendorff's Test Take 2-3 ml of plant extract in a test tube. Add few drops of dil.HCL and warm gently. Add Dragendorff's reagent	Reddish-brown precipitate	Alkaloid is present.
Wagner's Test Take 2-3 ml of plant extract in a test tube. Add few drops of dil.HCL and warm gently. Add Wagner's reagent	Reddish-brown precipitate	Alkaloid is present.

Table No.6 Result of Alkaloid test

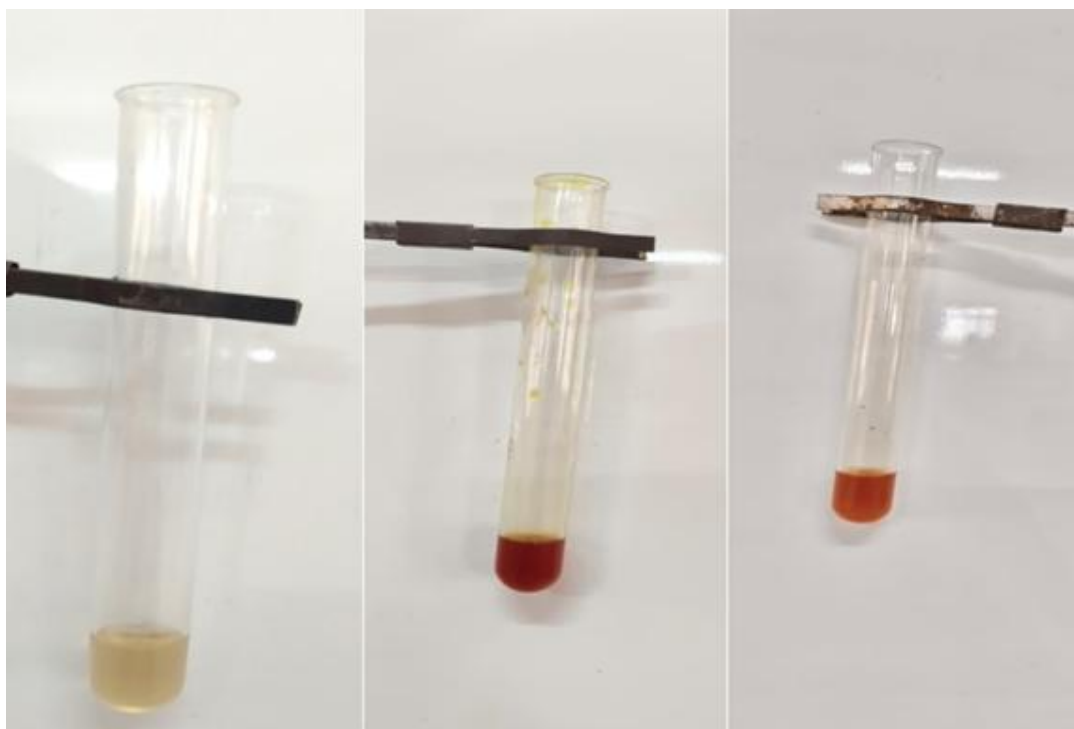


Fig.no.5 Result of Alkaloid test

**Flavonoids Test**

Shinoda Test Pink colour are observed Falvonoid is present



Table.No.7 Result of Shinoda test

Shinoda Test	Pink colour are observed	Falvonoid is present
--------------	--------------------------	----------------------



Fig.no.6 Result of Shinoda test

Table.No.8 Result of Saponin test

Foam Test	Persistent foam (>1cmheight)	Saponin is Present.
-----------	------------------------------	---------------------



Fig.no.7 Result of Saponin test



Tannins Test

Tannis Test	Black color observed	Tannin is Present
-------------	----------------------	-------------------

Table.No.9 Result of Tannin test

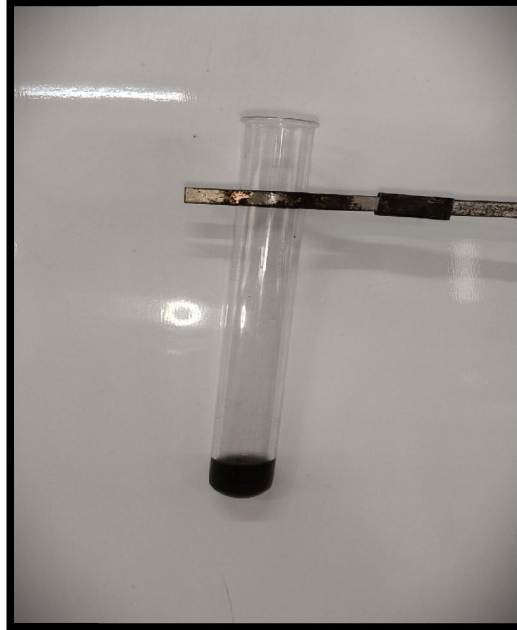


Fig.no.8 Result of Tannin test

Terpenoids Test

Salkowski Test	Brown ring present at junction of two chemicals.	Terpenoids is present
----------------	--	-----------------------

Table.No.10 Result of Terpenoids test

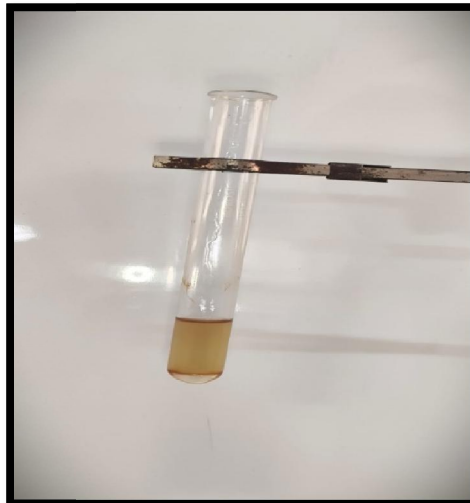


Fig.no.9 Result of Terpenoids test



**ASH VALUE**

Ash Value (% w/w) = 5%



Fig.no.10 Ash Value

**2.3 LOSS OF DRYING**

Loss on Drying (% w/w) =10%



Fig.no.11 Loss of Drying

**2.4 Anti-Glycation Assay By Titration Method:**

Observation Table:

Concentration (µg/mL)	Initial reading	Final reading	Titre value	Percentage Inhibition
0 (Blank)	0.0	10.0	10.0	-
20	0.0	9.5	9.5	5 %
40	0.0	8.7	8.7	13%
60	0.0	6.5	6.5	35%
80	0.0	6	6	40%
100	0.0	5	5	50%

Table No.11 Observation Table of Anti-Glycation Assay



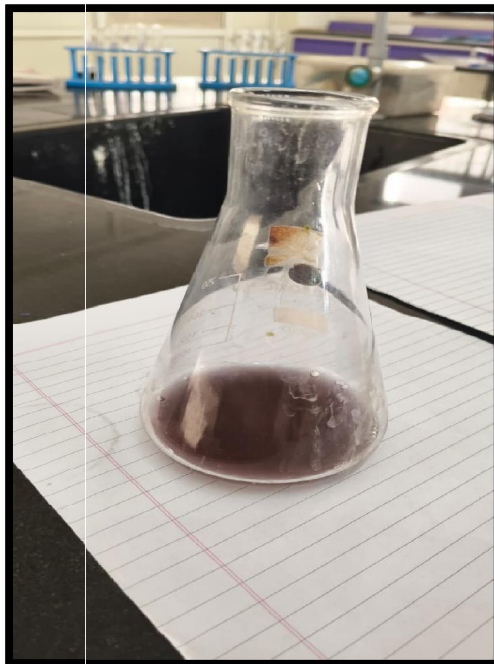


Fig.no.12 Result of Anti-Glycation Assay By Titration Method

## VI. CONCLUSION

In vitro antiglycation tests, physicochemical analysis, and phytochemical screening were used in this work to assess the antiglycation activity of Eucalyptus globulus leaf extract. The findings showed that because Eucalyptus globulus contains a variety of bioactive phytoconstituents, it has considerable therapeutic potential. Important secondary metabolites with antioxidant, anti-inflammatory, and medicinal qualities, including alkaloids, flavonoids, tannins, saponins, and terpenoids, were found by preliminary phytochemical screening. Advanced glycation end products (AGEs), which are linked to diabetes, aging, and a number of chronic illnesses, may be inhibited by these substances. The crude drug's acceptable purity and moisture content were confirmed by the physicochemical criteria, such as ash value (5%) and loss on drying (10%), which validated its suitability for experimental usage. Phytochemicals were effectively recovered from the leaves using the Soxhlet extraction technique with methanol or ethanol, yielding a concentrated crude extract appropriate for further study.

The titration approach of the antiglycation test demonstrated concentration-dependent glycation inhibition. Strong antiglycation action was demonstrated by the percentage inhibition rising from 5% to 50% as the extract concentration rose from 20  $\mu\text{g/mL}$  to 100  $\mu\text{g/mL}$ . Eucalyptus globulus may successfully lower glycation and AGE production, according to the maximal inhibition at 100  $\mu\text{g/mL}$ .

## REFERENCES

- 1] Trease and Evans Pharmacognosy. Evans WC. Trease and Evans Pharmacognosy. 16th ed. London: Elsevier; 2009.
- 2] Kokate's Practical Pharmacognosy. Kokate CK, Purohit AP, Gokhale SB. Practical Pharmacognosy. Pune: Nirali Prakashan; 2010.
- 3] Harborne's Phytochemical Methods. Harborne JB. Phytochemical Methods: A Guide to Modern Techniques of Plant Analysis. 3rd ed. London: Chapman & Hall; 1998.
- 4] Indian Materia Medica. Nadkarni KM. Indian Materia Medica. Mumbai: Popular Prakashan; 2005.



- 5] Textbook of Pharmacognosy and Phytochemistry. Shah B, Seth AK. Textbook of Pharmacognosy and Phytochemistry. New Delhi:Elsevier; 2010.
- 6] World Health Organization Monographs on Selected Medicinal Plants. World Health Organization. WHO Monographs on Selected Medicinal Plants. Geneva: WHO; 2002.
- 7] The Wealth of India: A Dictionary of Indian Raw Materials and Industrial Products. The Wealth of India: Raw Materials. New Delhi: CSIR; 2003.
- 8] Pharmacognosy. Khandelwal KR. Practical Pharmacognosy Techniques and Experiments. Pune: Nirali Prakashan; 2008.
- 9] Medicinal Plants of the World. Van Wyk BE, Wink M. Medicinal Plants of the World. Portland: Timber Press; 2004.
- 10] British Pharmacopoeia. British Pharmacopoeia. London: Her Majesty's Stationery Office; 2018.
- 11] Indian Pharmacopoeia. Indian Pharmacopoeia. Ghaziabad: Indian Pharmacopoeia Commission; 2018.
- 12] Phytochemistry. Sofowora A. Medicinal Plants and Traditional Medicine in Africa. Ibadan: Spectrum Books Ltd; 2008.
- 13] Eucalyptus Essential Oil Industry Review. Batish DR, Singh HP, Kohli RK, Kaur S. Eucalyptus essential oil as a natural pesticide and therapeutic agent. *Forest Ecology and Management*. 2008;256(12):2166–2174.
- 14] Advanced Glycation End Products Research. Singh R, Barden A, Mori T, Beilin L. Advanced glycation end-products: A review. *Diabetologia*. 2001;44(2):129–146.
- 15] Antioxidant Activity of Medicinal Plants. Rice-Evans C, Miller N, Paganga G. Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free Radical Biology and Medicine*. 1996;20(7):933–956.
- 16] Thin Layer Chromatography in Phytochemical Analysis. Wagner H, Bladt S. *Plant Drug Analysis: A Thin Layer Chromatography Atlas*. Berlin: Springer; 1996.
- 17] Phytochemical Screening Methods. Kokate CK. *Practical Pharmacognosy*. 4th ed. New Delhi: Vallabh Prakashan; 1994.
- 18] Medicinal Properties of Eucalyptus globulus. Santos SAO, Villaverde JJ, Silva CMS, Neto CP, Silvestre AJD. Chemical composition and biological activities of Eucalyptus globulus. *Industrial Crops and Products*. 2012;43:562–567.
- 19] Natural Antiglycation Agents. Peng X, Ma J, Chao J, Sun Z, Chang RC. Beneficial effects of natural compounds against glycation and AGE formation. *Journal of Ethnopharmacology*. 2011;137(1):1–13.
- 20] Medicinal Plant Phytochemicals. Cowan MM. Plant products as antimicrobial agents. *Clinical Microbiology Reviews*. 1999;12(4):564–582.

