

Formulation and Evaluation of Nitroglycerin Transdermal Film

**Shreya Govind Thombare, Sonaji Balu Farande, Snehal Narayan Bobade,
Nikita Sunil Kokate, Akshata A. Gosavi**

Sahakar Maharshi Kisanrao Varal Patil College of Pharmacy, Nighoj
shreyathombare03gmail.com

Abstract: *The objective of the present study was to formulate and evaluate transdermal films of Nitroglycerin using hydroxypropyl methylcellulose (HPMC), cellulose acetate phthalate, ethyl cellulose, and polypropylene glycol as rate-controlling membrane materials, and HPMC, cellulose acetate phthalate, and ethyl cellulose as drug-reservoir gels. Transdermal films of Nitroglycerin incorporating HPMC, cellulose acetate phthalate, and ethyl cellulose in varying blend ratios were prepared using the solvent-casting method.*

The thickness of the films ranged from 0.18 ± 0.02 mm to 0.22 ± 0.03 mm, indicating uniformity in film dimensions. The optimized formulation (NF1) demonstrated stability for a period of one to three months. All developed films were evaluated for drug content and in-vitro diffusion characteristics.

Keywords: *hydroxypropyl methylcellulose*

I. INTRODUCTION

In the present study, our objective was to develop transdermal therapeutic systems for the controlled delivery of the anti-anginal drug Nitroglycerin. Accordingly, investigations were undertaken on membrane-moderated therapeutic systems employing hydroxypropyl methylcellulose (HPMC), cellulose acetate phthalate, ethyl cellulose, and polypropylene glycol as rate-controlling membranes, while HPMC, cellulose acetate phthalate, and ethyl cellulose were used as drug-reservoir gels [1].

The use of such systems is particularly important for managing chronic conditions such as hypertension, which require long-term dosing to maintain therapeutic drug concentrations. Transdermal delivery of cardiovascular (CVS) drugs offers several advantages, including avoidance of hepatic first-pass metabolism and the ability to sustain constant plasma drug levels over an extended period, thereby reducing the frequency of dosing and improving patient compliance [2]. Transdermal drug delivery systems (TDDS), especially transdermal films, provide controlled release, improved bioavailability, and reduced dosing frequency. This review focuses on the theoretical aspects of materials used and methods adopted for the formulation and assessment of nitroglycerin transdermal films.

II. MATERIALS AND METHODS

Materials

Nitroglycerin was procured from Wockhardt Limited, Aurangabad. Hydroxypropyl methylcellulose (HPMC), cellulose acetate phthalate, ethyl cellulose, propylene glycol, and dimethyl sulfoxide were obtained from Frequent Pharmaceuticals Private Limited, Mumbai. All other excipients used in the study were of analytical grade (Table 1). Polymers determine the diffusion path length, affecting drug release kinetics. Plasticizers reduce intermolecular polymer interactions, increasing polymer chain mobility. [3]. The formulation and assessment of nitroglycerin transdermal film material involve several steps. First, the active pharmaceutical ingredient (nitroglycerin), along with excipients like polymers, plasticizers, and permeation enhancers, are selected and weighed. [16] The materials are then mixed, and the mixture is cast onto a backing film. After drying, the film is cut into the desired sizes. The assessment includes tests for physical properties (thickness, uniformity), drug content, in vitro drug release, and adhesion. [17] The film's performance is also evaluated in terms of its ability to deliver nitroglycerin through the skin, often using in vitro or in vivo studies. [18] The



goal is to create a transdermal film that effectively delivers nitroglycerin for therapeutic effects, usually for angina. First, the active pharmaceutical ingredient (nitroglycerin), along with excipients like polymers, plasticizers, and permeation enhancers, are selected and weighed. The materials are then mixed, and the mixture is cast onto a backing film. After drying, the film is cut into the desired sizes.[19]

The assessment includes tests for physical properties (thickness, uniformity), drug content, in vitro drug release, and adhesion. The film's performance is also evaluated in terms of its ability to deliver nitroglycerin through the skin, often using in vitro or in vivo studies. The goal is to create a transdermal film that effectively delivers nitroglycerin for therapeutic effects, usually for angina.[20] Nitroglycerin is a potent vasodilator used in the treatment of angina. Oral administration suffers from extensive first-pass metabolism, making the transdermal route a suitable alternative. Transdermal films provide controlled drug release, reduce dosing frequency, and minimize side effects.

FORMULATION OF NITROGLYCERIN TRANSDERMAL FILMS

1. Drug: Nitroglycerin.
2. Polymers: HPMC, PVP, Eudragit, Ethyl cellulose.
3. Plasticizers: PEG 400, Glycerin, Propylene glycol.
4. Solvents: Ethanol, Chloroform, Isopropyl alcohol.
5. Permeation Enhancers: Oleic acid, Menthol, Tween 80.

METHODS OF FILM PREPARATION

1. Solvent Casting Method.
2. Hot Melt Extrusion.
3. Adhesive Dispersion Method.

EVALUATION PARAMETERS

1. Thickness and Weight Variation
2. Folding Endurance
3. Tensile Strength
4. Moisture Absorption and Loss
5. Drug Content Uniformity
6. In Vitro Drug Release
7. Ex Vivo Skin Permeation Studies
8. FTIR, DSC, and XRD Compatibility Studies

MECHANISM OF DRUG RELEASE

Diffusion-controlled release through polymer matrix and enhanced permeation through skin layers.

Procedure

A beaker containing 15 mL of purified water and three drops of glycerin was stirred for 15 minutes. HPMC and PVP were then added gradually with continuous stirring for 30 minutes [4]. The drug and propylene glycol were incorporated into the above mixture, and the resulting mass was poured into a transdermal mold. The films were dried in a hot air oven at 50 °C for 15 minutes and subsequently stored in desiccators (Table 5).

Evaluation of Transdermal Films

Physical Appearance

All formulated films were visually examined for color, flexibility, homogeneity, and surface smoothness [5].

Thickness

Film thickness was measured using a screw gauge (micrometer) [6].

Weight Variation

Each patch was weighed individually, and the average film weight was calculated [7].



Moisture Content

Films (n = 3) were individually weighed and stored in a desiccator containing calcium chloride at 37 °C for 24 hours [8]. Moisture content was determined from the difference between initial and final weights.

Flatness

Variation in film length due to non-uniform flatness was assessed by measuring the shrinking of strips [9].

Tensile Strength

Tensile strength was determined by gradually adding weights to the pan to increase the pulling force until the patch ruptured [10].

Folding Endurance

Folding endurance was measured by repeatedly folding a film at the same point until it broke. The number of folds tolerated before breaking represented the folding endurance value [11].

Water Vapour Transmission Rate (WVTR)

The film was secured over the opening of a glass vial containing 3 g of fused calcium chloride (desiccant) using adhesive tape. The vial was weighed and stored in a desiccator containing a saturated potassium chloride solution to maintain 84% relative humidity. The vial was reweighed at 24-hour intervals over 72 hours [12].

Drug Content

A 1 cm² film sample was placed in a beaker containing 100 mL of phosphate buffer (pH 7.4) and stirred with a Teflon-coated magnetic bead for 5 hours [5].

In Vitro Drug Permeation Studies

In vitro release studies were carried out using a modified Franz diffusion cell for 12 hours. At predetermined intervals, aliquots were withdrawn from the receptor compartment and analyzed using an appropriate analytical method [13].

Stability Studies

Stability testing of Nitroglycerin transdermal films was conducted at 40 °C/75% RH for one and three months. Films were evaluated for in-vitro diffusion characteristics at each interval [15].

III. RESULTS AND DISCUSSION

Compatibility Studies

The FTIR spectra of pure Nitroglycerin are presented in Figures 1–5.

Evaluation of Nitroglycerin Transdermal Films

Physicochemical Properties

The formulated films were thin, flexible, elastic, smooth, and transparent or translucent (Table 2).

Thickness

Film thickness ranged from 0.18 ± 0.02 mm to 0.22 ± 0.03 mm, indicating uniformity (Table 2).

Weight Variation

Weights of the patches varied between 150 ± 2.2 mg and 221 ± 2.6 mg, demonstrating consistency across batches (Table 2).

Moisture Content

Moisture content ranged between 2.432 ± 0.01% and 3.354 ± 0.02% (Table 2).

Ingredients	NF1	NF2	NF3	NF4	NF5	NF6
Nitroglycerin [mg]	100	100	100	100	100	100
Hydroxy Propyl Methyl Cellulose[Mg]	400	-	-	100	100	-
Cellulose Acetate phthalate [mg]	-	400	-	200	-	100
Ethyl cellulose[mg]	-	-	400	-	200	200
Propylene glycol [W/V]	40%	40%	40%	40%	40%	40%
Alcohol[ml]	10	10	10	10	10	10
Dimethylsulfoxide	20%	20%	20%	40%	40%	40%

Table 1 : Ingredients of Transdermal Films



Table 2: Evaluation of Transdermal Films

Parameters	NF1	NF2	NF3	NF4	NF5	NF6
Physical Appearance	Satisfactory	Satisfactory	Satisfactory	Satisfactory	Satisfactory	Satisfactory
Thickness (mm)	0.20 ± 0.04	0.18 ± 0.02	0.19 ± 0.03	0.22 ± 0.03	0.20 ± 0.04	0.21 ± 0.05
Weight variation (mg)	210 ± 1.19	170 ± 2.20	150 ± 2.2	221 ± 2.6	202 ± 5.0	187 ± 2.2
Moisture content	3.113 ± 0.04	3.321 ± 0.31	3.354 ± 0.02	3.131 ± 0.03	2.432 ± 0.01	3.114 ± 0.23
Flatness (I)	98%	98%	98%	98%	98%	98%
Tensile strength (Nm ⁻²)	11.12 ± 1.11	12.14 ± 1.31	11.89 ± 1.57	12.12 ± 1.31	12.23 ± 1.78	10.21 ± 3.00
Folding endurance (F)	170.1 ± 3.10	150 ± 2.34	160 ± 1.10	134 ± 5.3	143.2 ± 4.5	100 ± 2.43
WVTR (g/m ²)	3.110 ± 0.24	3.232 ± 0.32	3.121 ± 0.14	2.189 ± 0.04	2.51 ± 0.34	2.01 ± 0.21
Drug Content (%)	98.2 ± 0.2	97.1 ± 0.1	96.9 ± 0.2	97.8 ± 0.3	96.3 ± 0.2	97.2 ± 0.3

Time(Min)	NF1	NF2	% of Drug release NF3	NF4	NF5	NF6
5	1.54	4.06	5.96	11.68	5.58	13.14
10	9.62	10.96	11.24	22.36	19.42	26.98
15	15.96	21.46	23.59	34.64	28.32	33.16
20	20.68	31.58	44.64	49.32	33.16	43.94
25	31.58	45.42	59.32	53.16	47.06	56.31
30	45.42	57.06	60.90	65.32	57.08	68.64
35	57.06	64.80	74.80	78.64	79.42	70.98
40	80.90	82.48	81.00	84.90	82.64	81.10
45	97.34	83.22	92.12	95.86	91.26	93.56

Table 3:- Cumulative % Drug Release of Nitroglycerin Transdermal Films

Time(Min)	Standard	Cumulative%Drug Release After 1 Month
5	1.54	1.59
10	9.62	9.79
15	15.96	16.00
20	20.68	22.68
25	31.58	33.58
30	45.42	46.42
35	57.06	58.06
40	80.90	81.90
45	97.35	97.37

Table 4: Stability studies of the optimized formulation NF1

Drug Content (%)	After 1 Month
98.2±0.2	98.3±0.1

Table 5: Stability studies of the Drug Content of optimized formulation N



Flatness

Flatness measurements indicated that all formulations exhibited approximately 98% flatness, demonstrating minimal dimensional deviation (Table 2).

Tensile Strength

Formulations NF1 to NF6 showed tensile strength values ranging from 11.12 ± 1.11 to 12.23 ± 1.31 , indicating satisfactory mechanical resistance and cohesive film characteristics (Table 2).

Folding Endurance

Folding endurance values for formulations NF1 to NF6 ranged from 100 ± 2.43 to 170.1 ± 3.10 , reflecting good flexibility and structural integrity (Table 2).

Water Vapour Transmission Rate (WVTR)

The WVTR values for formulations NF1 to NF6 ranged from 2.01 ± 0.21 to 3.232 ± 0.32 , indicating acceptable moisture permeability characteristics (Table 2).

Drug Content

The drug content of the films was found to be between $96.3 \pm 0.2\%$ and $98.2 \pm 0.2\%$, demonstrating uniform drug distribution within the formulations (Table 6).

In-Vitro Drug Permeation Studies

Formulation NF1 exhibited the highest drug release, with 97.34% of Nitroglycerin released during the study period (Figure 6; Table 3).

In-Vitro Drug Release Kinetics

The in-vitro drug diffusion data were fitted to various kinetic models to evaluate the mechanism of release (Table 4).

Stability Study

Formulation NF1 was subjected to stability testing for one to three months. The films were evaluated for drug content and in-vitro diffusion during the stability period (Table 5; Figures 6 and 7).

IV. CONCLUSION

The results of the present study demonstrate that the Nitroglycerin-loaded transdermal films follow first-order release kinetics. The optimized formulation, NF1, remained stable for one to three months and retained acceptable drug content and in-vitro diffusion characteristics. These findings indicate that the limitations associated with oral administration of Nitroglycerin—such as dissolution rate-limited absorption and gastric side effects—can be effectively minimized by administering the drug transdermally through polymeric films. Nitroglycerin transdermal films provide a promising alternative to conventional dosage forms. Optimized formulations ensure sustained release, improved bioavailability, and enhanced therapeutic efficacy.

REFERENCES

- [1] R R Thenge, K G Mahajan, H S Sawarkar, V S Adhao, and P S Gangane. Formulation and evaluation of transdermal drug delivery system for lercanidipine hydrochloride. International Journal of PharmTech Research, 2(1):253–258,2010.
- [2] Y Ramesh, A K M Anjana, D B Manjula, K San-keerthana, L P Sri, and A Vasanthi. Formulation and evaluation of atenolol transdermal patches. Creative Journal of Pharmaceutical Research, 1(2):55–65, 2015.
- [3] P Koteswararao, S Duraivel, K P SampathKumar, and Debjit Bhowmik. Formulation And Evaluation Of Transdermal Patches Of Anti-Hypertensive Drug Metoprolol Succinate. Indian Journal of Research in Pharmacy and Biotechnology, 1(5):629–639, 2013.
- [4] N Divya, R Hemalatha, M Nirosha, and S Ramkanth. Fabrication and evaluation of transdermal matrix patches of Metoprolol Tartrate. International Research Journal of Pharmaceutical and Applied Sciences, 7(4):31– 35, 2017.
- [5] K S Vijayakumar, S Parthiban, G P Senthilkumar, and T Tamiz Mani. Formulation And Evaluation of Gliclazide Loaded Ethosomes As Transdermal Drug Delivery Carriers. Asian Journal of Research in Biological and Pharmaceutical Sciences, 2(2):89–98, 2014.



- [6] T Li, C Ren, M Wang, L Zhao, X Wang, and L Fang. Optimized preparation and evaluation of indomethacin transdermal patch. *Asian Journal of Pharmaceutical Sciences*, 2(6):249–259, 2007.
- [7] Yerikala Ramesh and Vadhireddy Sireesha. Transdermal patch of ramipril loaded chitosan nanoparticles dispersed in carbopol gel. *Journal of Drug Delivery and Therapeutics*, 7(6):56–65, 2017.
- [8] D. N. Reddy. Design, development and characterization of clopidogrel bisulfate transdermal drug delivery system. *Asian Journal of Pharmaceutical and Clinical Research*, (8):277–280, 2015.
- [9] S Sucharitha and CH. Praveen Kumar. Ethosomes - a novel vesicular transdermal drug carrier. *International Journal of Pharmacometrics and Integrated Biosciences*, 1(1):1–6, 2016.
- [10] Shaila Lewis, S Pandey, and N Udupa. Design and evaluation of matrix type and membrane controlled transdermal delivery systems of nicotine suitable for use in smoking cessation. *Indian Journal of Pharmaceutical Sciences*, 68(2):179–184, 2006.
- [11] P Anitha, S Ramkanth, M T S Saleem, K Umasankari, B P Reddy, and M Chetty. Preparation, in-vitro and in-vivo characterization of transdermal patch containing glibenclamide and atenolol: a combinational approach. *Pakistan Journal of Pharmaceutical Sciences*, 24(2):155–163, 2011.
- [12] R M Viswanatha, R V Jayashankar, Y Ramesh, and I Venkateswarlu. Formulation and evaluation of fluconazole transdermal patches. *International Journal of Institutional Pharmacy and Life Sciences*, 1:18–29, 2011.
- [13] M Aqil and Asgar Ali. Monolithic matrix type transdermal drug delivery systems of pinacidil monohydrate: in vitro characterisation. *European Journal of Pharmaceutics and Biopharmaceutics*, 54(2):161–164, 2002.
- [14] Christopher W Jeans and Charles M Heard. A therapeutic dose of primaquine can be delivered across excised human skin from simple transdermal patches. *International Journal of Pharmaceutics*, 189(1):1–6, 1999.
- [15] Y. S Rhee, S. Y Kwon, C. W Park, N. Y Choi, W. J Byun, S. C Chi, and E. S Park. Characterization of monolithic matrix patch system containing tulobuterol. *Archives of Pharmacological Research*, 31(8):1029–1034, 2008.
- [16] Guy, R. H., & Hadgraft, J. (2003). *Transdermal Drug Delivery*. Marcel Dekker, New York.
- [17] Baker, R. W. (1987). *Controlled Release of Biologically Active Agents*. Wiley-Interscience.
- [18] Kalia, Y. N., et al. (2004). “Principles of skin permeation and formulation strategies for transdermal delivery.” *Advanced Drug Delivery Reviews*, 56(5), 627–648.
- [19] Basu, B., et al. (2010). “Formulation and evaluation of transdermal films of nitrates.” *International Journal of Pharmaceutical Sciences Review and Research*, 4(2), 79–84.
- [20] Margetts, L., & Sawyer, R. (2007). “Transdermal drug delivery: principles and opioid therapy.” *Continuing Education in Anaesthesia Critical Care & Pain*, 7(5), 171–176.

