

Synthesis, Characterization of Magnetic Iron Oxide Nanoparticles and its Toxicity Assessment in Tilapia fingerlings

Nutan Rathod

Head, Associate Professor, PG Department of Zoology
Moolji Jaitha (Autonomous) College, Jalgaon, Maharashtra, India

Abstract: Nowadays, the most recent advances in nanoscience and nanotechnology are widely used in a variety of industries, including pharmaceuticals, medicine, electronics, robotics, and tissue engineering. The current study focuses on the chemical synthesis of Magnetic Iron Oxide Nanoparticles, Followed by characterization using UV-visible spectrometry, FTIR, SEM, and EDS. Toxicity tests were performed on fresh water fish Tilapia (*Oreochromis mosambicus*) fingerlings exposed to as synthesized Magnetic Fe_3O_4 NPs LC_{50} concentration of Fe_3O_4 NPs were 12.45 mg/L for 96 hrs and sub-lethal LC_{10} concentrations (10 ppm and 20 ppm) for 96 hours. When compared to controls, treated groups showed significant changes in biochemical indicators such as protein and catalase activity after 96 hours, which may be related to Fe_3O_4 NPs in fish muscles. The cause of nanoparticle toxicity is attributed to their own specific characteristics of greater surface-to-volume ratio, chemical composition, size, and dosage, body retention, and organ specific toxicity. The comprehensive data can provide the important empirical parameters as well as recent field developments. The extensive data is able to provide both the crucial empirical aspects and the most recent advancements in this field.

Keywords: Fe_3O_4 NPs, Characterization, toxicity, Fish and Biochemical test

I. INTRODUCTION

Iron Oxide Nanoparticles (NPs) have earned a significant attention at present, in relation to their unique properties which has basic scientific value but also for a wide range of biomedical and technological applications [1] [2]. The synthesis of Iron Oxide Nanoparticles (NPs) involves a range of intricate, costly, physical, and chemical procedures that result in dangerous, toxic waste that is detrimental to human health and the environment [3]. The synthesis of Iron Oxide Nanoparticles (NPs) involves a range of intricate, costly, physical, and chemical procedures that result in dangerous, toxic waste that is detrimental to human health and the environment. Nanoparticle research is expanding due to their unique physical and chemical properties [4] [5] [6]. The high surface-area-to-volume ratio of nanoparticles opens up numerous opportunities for producing antibacterial medicines to treat bacterial illnesses [7]. The possible health concerns posed to non-target creatures in the environment make it critical to set adequate rules for safe IONP management and disposal in order to protect the aquatic environment [8] [9]. However, despite the fact that it has been shown that Iron Oxide Nanoparticles can enter the human body through a number of routes, such as ingestion, injection, dermal penetration, and inhalation, little is known about the impact of these particles on human health, their potential side effects, and how their systematic circulation can change them in different tissues [10] and control over the production, use, and discharge of these nanoparticles in aquatic environments. When compared to conventional formulations, the growing exploration of NPs revealed many properties such as enhanced magnetic, catalytic, optical, and mechanical properties. Many commercial applications, such as fillers, catalysts, filters, semiconductors, microelectronics, and so forth, use these materials extensively, exposing people both directly and indirectly to environmental and medical risks. According to investigator [11], there are certain nanoparticles that are used in biomedical applications like protein immobilization, magnetic resonance imaging (MRI), gene and medication delivery, cell separation, and so forth. The persistence of nanoparticles after long-term exposure to Fe_3O_4 NPs through treatment



withdrawal to confirm the persistent impact of nanoparticles on the exposed organism were investigated [12]. Iron nanoparticles (FeNPs), showing, compete favourably with dangerous chemical pollutants in water and the environment [13].

Although magnetic nanoparticles must have specific properties, such as a persistent, switchable magnetic state in data storage applications to indicate bits of information that are influenced by thermal effects [14] and other investigator evaluated the altered the quantity of Iron Oxide Nanoparticles and noted the changes on growth, haematological and biochemical parameters of Koi Carp [15].

Furthermore, the potential eco-toxicological consequences, bioavailability, and detrimental effects of nanoscale contaminants in the aquatic environment can be evaluated using appropriate biomarkers for aquatic species, like fish [16]. Therefore, it is necessary to synthesize a simple, cost-effective, reproducible technique to synthesize Fe_3O_4 nanoparticles. The toxicity of MNPs is thus determined by a variety of factors, including size, shape, structure, surface modification, concentration, dosage, bio-distribution, bioavailability, solubility, immunogenicity, and pharmacokinetics, making it critical to understand such variables [17] [18]. The precipitation technique is used to chemically method for nanoparticles and Tilapia was selected for this study since it is a traditional nutritious eating fish in India. Hence in this present work is link up for synthesis, characterization, and toxicological assessment of Iron Oxide nanoparticles (Fe_3O_4) in fish Tilapia fingerlings.

II. MATERIAL AND METHODS

The chemicals were all of analytical grade. The precipitation method for FeCl_3 and FeSO_4 was used for synthesis of Iron Oxide Nanoparticles and NaOH were used as precursors [19] [20]. Characterization of Iron Oxide NPs carried out by FTIR, SEM, EDS, and UV-visible spectrometry. Characterizations (SEM and EDS) of Iron Oxide NPs were carried out at Physic Department of SPP University, Pune and also FTIR at Chemical Technology department of KBC NM University Jalgaon. Healthy Tilapia fingerlings were procured from Fish Farm in Jalgaon, Maharashtra, India, and acclimatized for about 15 days before the experiment began. For assessment of Fe_3O_4 NP toxicity, fish with an average length of 12 ± 0.5 cm and average weight of 10.2 ± 4 g were selected. Toxicity tests were conducted by synthesized Fe_3O_4 NPs at exponential concentrations (10 ppm and 20 ppm) for 96 hrs. Significant differences in biochemical parameters protein and catalase activity were found after 96 hours in treated groups compared to controls, which could be attributed to Iron Oxide NPs in fish muscles. The estimation of protein concentration was done by Biuret method using kit. The technique was used to estimate Catalase enzyme activity [21].

III. RESULT AND DISCUSSION

Synthesized Iron oxide nanoparticles by chemical method as NaOH is added to FeCl_3 and FeSO_4 , the colour of solution changes as of reddish brown to black, is presented in Fig.1 depicts the formation of Iron Oxide Nanoparticles (Fe_3O_4 NPs). Precipitation was observed at pH to 11. Fig 1. Indicates the experimental design. Fig 2. Chemical Synthesis of Iron Oxide Nanoparticles (Fe_3O_4 NPs). The characterization of Iron Oxide nanostructures was further confirmed by the UV-Vis spectra, FTIR, SEM, EDS and (Figure 3, 4, 5 and 6) signifying high density chemically synthesized Fe_3O_4 NPs.

Characterization of Iron Oxide NPs

UV-Vis Spectrophotometry Analysis

Preliminary confirmation was obtained by measuring the absorbance of the produced iron oxide nanoparticles in UV/Vis spectra within the 200–800 nm range. As shown in the present study, the concentrations of the associated functional groups may be directly correlated with the band of absorption intensities. The synthesis of iron oxide nanostructures was further confirmed by the UV-Vis spectra (Figure 2), which displayed high density chemically generated Fe_3O_4 . The findings are consistent with earlier research [22].

Fourier transform infrared (FTIR) spectra of Iron Oxide Nanoparticles

FTIR Functional Group bond of Fe_3O_4 NPs. Fourier Transform Infrared spectroscopy studies were performed to identify the probable functional groups responsible for the reduction of Fe ions by chemically synthesized iron oxide



nanoparticles. FTIR characterization of Fe_3O_4 nanoparticles were carried out shown in Figure 4 and Table 1, the data plot indicates the wave number of infrared light as prominent absorption peaks in particular wavenumbers that result from the vibration of certain functional groups. FTIR is an analytical technique used to identify the functional groups in a sample as well as determine the microcrystalline structure of the particles. The 4000–500 cm^{-1} range was studied using FTIR spectroscopy, which is sensitive to molecule structure, conformation, and environment. In Figure 3, the FTIR spectra reveal the three characteristic peaks at 670 cm^{-1} , and 855 cm^{-1} which are attributed to the Fe-O bond [23].

FE-SEM Images and EDS

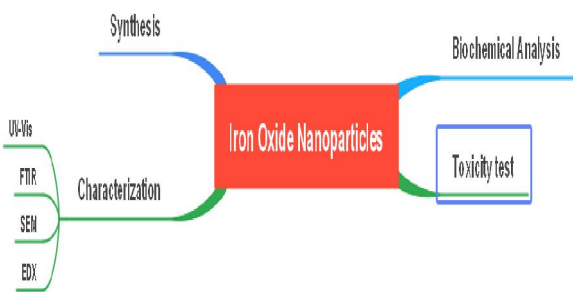
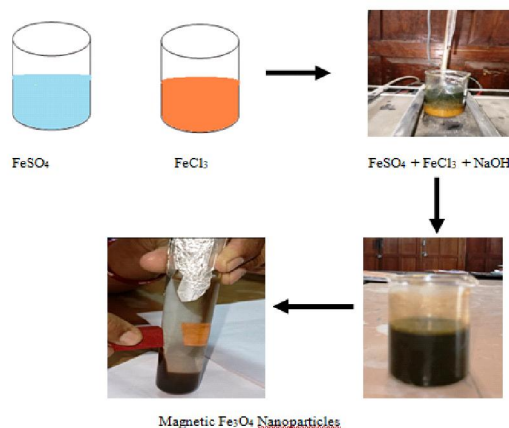
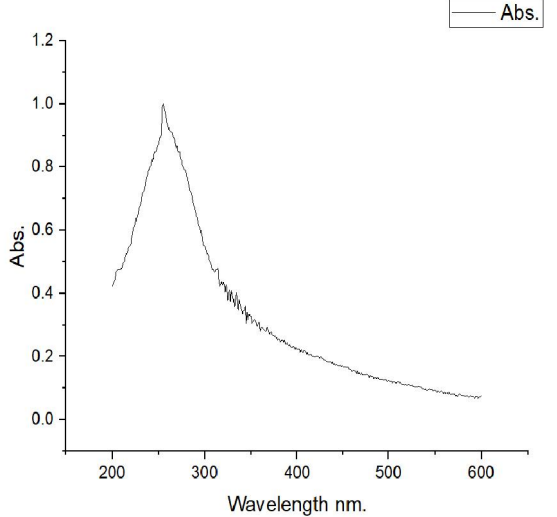
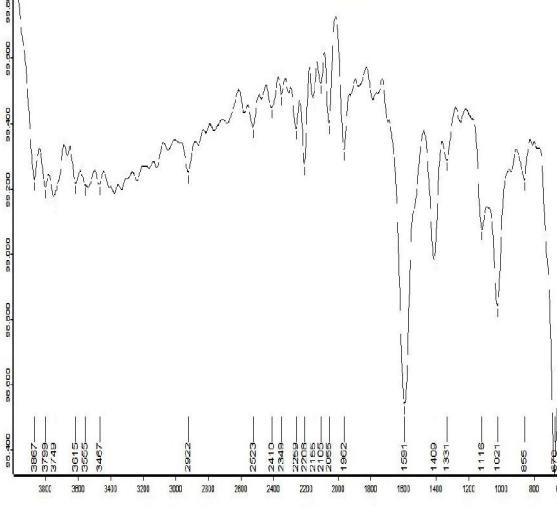
SEM image of Fe_3O_4 , the image depicts Fe_3O_4 particles formed of small particles. The FE-SEM examination reveals information about the surface shape and particle size of the produced samples. Figure 4 displays the iron oxide nanoparticle FE-SEM analysis. Spherical particles of different sizes and dispersion can be seen in microscopic pictures. Furthermore, it was shown that the ratio of particle agglomeration increased due to the electrostatic contact between the layers of nanoparticles. Similar results noted from SEM analysis of Iron Oxide Nanoparticles [24] [25].

Toxicity and Biochemical analysis

Toxicity tests were performed on fresh water fish Tilapia (*Oreochromis mosambicus*) fingerlings exposed to as synthesized Magnetic Fe_3O_4 NPs at LC_{50} concentration of Fe_3O_4 NPs were 12.45 mg/L for 96 hrs, and sub-lethal LC_{10} concentrations (10 ppm and 20 ppm) for 96 hours. Preliminary toxicity analyses have been carried out on Tilapia fingerlings to assess the 96 hrs median lethal concentration of Fe_3O_4 . Significant variations in biochemical parameters, including protein and catalase activity, were seen after 96 hours between the treatment and control groups. Effect of Iron Oxide Nanoparticles on the Catalase (CAT) in Gills and Muscles of the fish Tilapia ($\mu\text{mol H}_2\text{O}_2/\text{mg protein}/\text{min}$) noted in Table 3. Whereas Effect of Iron Oxide Nanoparticles on the Catalase (CAT) in Gills and Muscles of the fish Tilapia ($\mu\text{mol H}_2\text{O}_2/\text{mg protein}/\text{min}$) showed in Graph 1. Similar results are noted by [26] [27]. The non-uniform scattered nanoparticles morphology was noted, as seen in Fig 5. Iron oxide nanoparticles synthesized chemically are hexagonal and spherical in structure. In earlier studies on the acute toxicity of nanometals to aquatic life, concentrations in the mg–lg/L range can be fatal for fish. These results indicate that iron oxide nanoparticles prefer to combine due to intrinsic magnetic attraction [28]. Fish tissues all showed a marked reduction in protein levels following exposure to iron oxide nanoparticles. Acute toxic stress and generalized increased proteolytic activity lead protein breakdown to outweigh synthesis, suggesting a tissue state that favours anaerobic respiration to meet energy demands when aerobic oxidation is diminished [4] [29] [30] [31]. Earlier investigation was identified fish mortality in higher amounts of Iron Oxide NP treated groups, which could have resulted from an excessive accumulation of Iron Oxide NPs in the fish's body [10] [16]. The findings are the same as those of former research conducted [32]. Several publications have investigated Magnetic Iron Oxide Nanoparticles [33] [34] [35] [36] based on their biological activity, which includes antibacterial, toxicity, and drug delivery, they have been employed as model organisms in bio-system multifunctional nanomaterials. The biological system adjusts the activities of antioxidants like catalase (CAT) and related enzymes as ROS levels rise in order to build a first line defence mechanism [37]. Thus, the increase in catalase activity implies that iron oxide nanoparticles (NPs) have the ability to cause oxidative stress in fish [38] [39] [40] [41].



TABLE I

	
<p>Fig 1. Experimental design</p>	<p>Fig 2. Chemical Synthesis of Iron Oxide Nanoparticles (Fe₃O₄ NPs)</p>
	
<p>Fig.3 : UV of Fe₃O₄ NPs</p>	<p>Fig. 4 FTIR of Fe₃O₄ NPs</p>



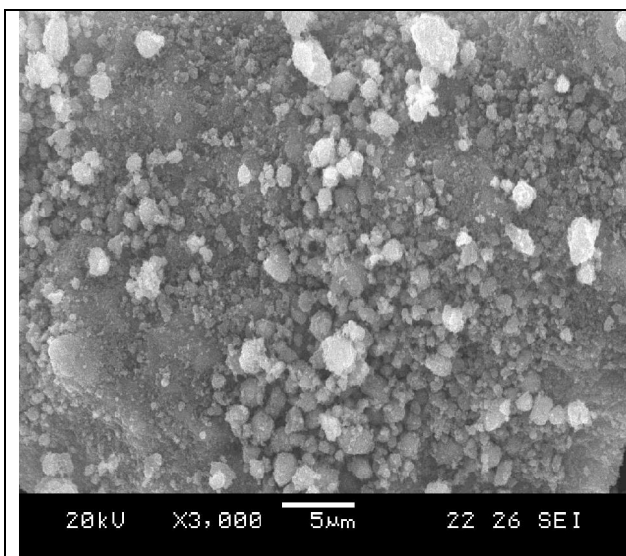


Fig 5. SEM Image of Iron oxide Nanoparticles

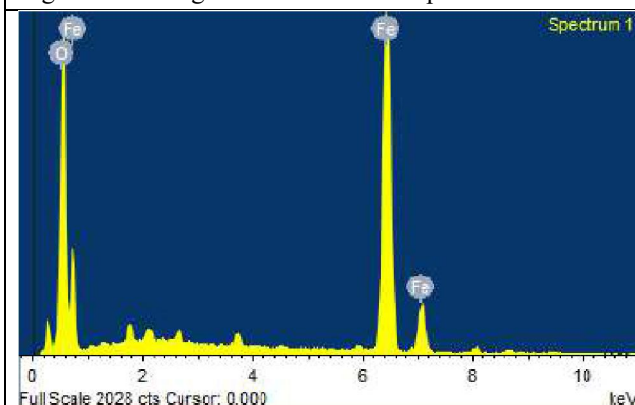


Fig 6. EDS Image of Iron oxide Nanoparticles

Sr. No.	Wave Number (cm-1) Fe ₃ O ₄ NPs	Functional Group
1	658-506	Fe-O stretching
2	1672-1367	H-O-H bending
3	3028-3400	O-H stretching
4	2920-2855	C-H stretching

Table 1. FTIR Functional Group bond of Fe₃O₄ NPs

Element	Weight %	Atomic %
O K	44.03	73.30
Fe K	55.97	26.70
Total	100.00	

Table 2. EDS area analysis of Iron Oxide Nanoparticle

Organs	Control	Treated	
		10 ppm	20 ppm
Gills	14.24±0.2	21.24±0.2*	18.6±0.4*
Muscles	34.22±1.22	40.32±2.12***	43.8±2.45*

P < 0.05; ** = P < 0.01; *** = P < 0.001

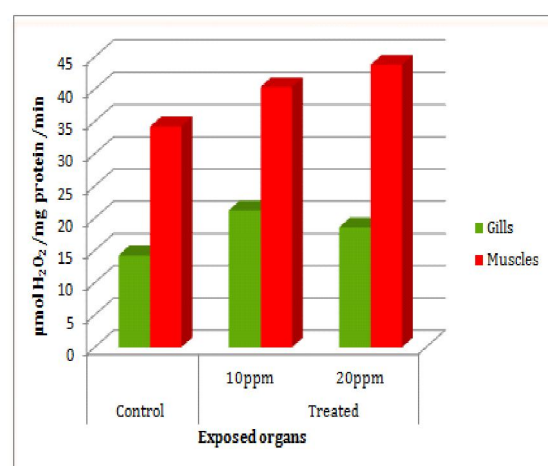


Table 3. Effect of Iron Oxide Nanoparticles on the Catalase (CAT) in Gills and Muscles of the fish Tilapia (µmol H₂O₂ / mg protein / min)

Graph 1. Effect of Iron Oxide Nanoparticles on the Catalase (CAT) in Gills and Muscles of the fish Tilapia (µmol H₂O₂ / mg protein / min)



IV. CONCLUSION

The safety assessment of synthetic Iron Oxide NPs included extensive characterisation, and fish toxicity testing, resulting in a better knowledge of their possible biological affects and environmental hazards. Using methods like UV-visible spectrometry, FT-IR, FESEM, EDS the physical and chemical characteristics of Iron Oxide NPs were fully examined. Present investigations showed that nanoparticles' size and form have a big impact on biological systems. Present data observed acute toxicity observed at LC50 concentration of Fe₃O₄ NPs were 12.45 mg/L for 96 hrs and toxicity at 10 ppm and 20 ppm for 96 hours freshwater fish *Tilapia* species exposed to as synthesized Fe₃O₄ NPs at LC10 concentration. After 96 hours, there were significant differences in biochemical measures such as protein and catalase activity in treated groups compared to controls, which could be due to iron oxide NPs in fish muscles. The concentration used in this study had a significant impact catalase activity on muscles of *Tilapia* fingerlings. These parameters are potentially useful as biomarkers or biological end points for determining the toxicity of manufactured Nanoparticles in aquatic creatures. The current study's findings emphasize the importance of safe metal oxide disposal and practices. However, further research is needed on the direct impact of metal ions and/or the release of metal ions from Nanoparticles.

V. ACKNOWLEDGMENT

We are greatly acknowledged to Principal Moolji Jaitha (Autonomous) College, Jalgaon for providing research facilities and also KBC North Maharashtra University, Jalgaon (M.S., India) for their financial support under VCRM Scheme.

REFERENCES

- [1]. Sophie Laurent, Delphine Forge, Marc Port, Alain Roch, Caroline Robic, Luce Vander Elst, Robert N Muller (2008). Magnetic iron oxide nanoparticles: synthesis, stabilization, vectorization, physicochemical characterizations, and biological applications, *Chem. Rev.*, 108, 2064–2110. <https://doi.org/10.1021/cr068445e>.
- [2]. Campos, E. A., Pinto, D. V. B. S., Oliveira, J. I. S. D., Mattos, E. D. C., & Dutra, R. D. C. L. (2015). Synthesis, characterization and applications of iron oxide Nanoparticles-a short review. *Journal of Aerospace Technology and Management*, 7, 267-276. <https://doi.org/10.5028/jatm.v7i3.471>.
- [3]. Klaine, S. J., Koelmans, A. A., Horne, N., Carley, S., Handy, R. D., Kapustka, L., Nowack, B., & von der Kammer, F. (2012). Paradigms to assess the environmental impact of manufactured nanomaterials. *Environmental Toxicology and Chemistry*, 31(1), 3-14. <https://doi.org/10.1002/etc.733>.
- [4]. Brigitta Szalay (2012). Iron oxide Nanoparticles and their toxicological effects: in vivo and in vitro studies., PhD Thesis., University of Szeged.
- [5]. Ali, A., Zafar, H., Zia, M., ul Haq, I., Phull, A. R., Ali, J. S., & Hussain, A. (2016). Synthesis, characterization, applications, and challenges of iron oxide nanoparticles. *Nanotechnology, science and applications*, 49-67. <https://doi.org/10.2147/NSA.S99986>
- [6]. Mashjoor, S., Yousefzadi, M., Zolgharnain, H., Kamrani, E., & Alishahi, M. (2018). Organic and inorganic nano-Fe₃O₄: *Alga Ulva flexuosa*-based synthesis, antimicrobial effects and acute toxicity to briny water rotifer *Brachionus rotundiformis*. *Environmental Pollution*, 237, 50-64. <https://doi.org/10.1016/j.envpol.2018.02.036>.
- [7]. Diez-Pascual, A. M. (2020). Recent Progress in Antimicrobial Nanomaterials, *Nanomaterials (Basel)*, 10 (11), 2315. <https://doi.org/10.3390/nano10112315>.
- [8]. Puthan Variyam Vidya Balakrishnan, Goran Gajski, and Kumari Chidambaran Chitra (2024). The effects of iron oxide nanoparticles on antioxidant capacity and response to oxidative stress in *Mozambique tilapia*., *Arh Hig Rada Toksikol* 2024;75:125-136. <https://doi.org/10.2478/aiht-2024-75-3826>.
- [9]. Xuan L, Ju Z, Skonieczna M, Zhou P, Huang R. Nanoparticles-induced potential toxicity on human health: Applications, toxicity mechanisms, and evaluation models. *MedComm* 2023;4(4):e327. <https://doi.org/10.1002/mco2.327>.



- [10]. Keerthika, V., Ramesh, R., & Rajan, M. R. (2017). Toxicity assessment of iron oxide Nanoparticles in *Labeo rohita*. *International Journal of Fisheries and Aquatic Studies*, 5(4), 01-06.
- [11]. Sun, W., Mignani, S., Shen, M. and Shi, X. (2016): Dendrimer-based magnetic Iron Oxide nanoparticles: their synthesis and biomedical applications. *Drug Discovery Today*. 21(12): 1873–1885. <https://doi.org/10.1016/j.drudis.2016.06.028>.
- [12]. Vidya P.V. and Chitra K.C. (2019): Irreversible Histopathological Modifications Induced by Iron Oxide Nanoparticles in the Fish, *Oreochromis mossambicus* (Peters, 1852)., *Biological Forum – An International Journal* 11(1): 01-06.
- [13]. Takai ZI, Mustafa MK, Asman S. Synthesis and characterization of magnetite (Fe₃O₄) nanoparticles with different levels of aniline dimer-COOH by Co-precipitation method. *Nanosci Nanotech: Nano-SciTech*. 2019. <https://doi.org/10.1063/1.5124670>.
- [14]. Abolfazl Akbarzadeh, Mohamad Samiei and Soodabeh Davaran (2012). Magnetic nanoparticles: preparation, physical properties, and applications in biomedicine., *Nanoscale Research Letters*, 7:144. <http://www.nanoscalereslett.com/content/7/1/144>
- [15]. Sangeetha K., Rajan M.R. (2021). Evaluation of Different Quantity of Iron Oxide Nanoparticles on Growth, Haematological and Biochemical Characteristics of Koi Carp . *Agricultural Science Digest*. 41(2): 338-344. doi: 10.18805/ag.D-5236.
- [16]. Remya, A.S.; Ramesh, M.; Saravanan, M.; Poopal, R.K.; Bharathi, S.; Nataraj, D (2015). Iron Oxide nanoparticles to an Indian Major Carp, *Labeo rohita*: Impacts on hematology, iono regulation and gill Na⁺/K⁺ATPase activity. *J. King Saud Univ. Sci.* 2015, 27, 151–160. <https://doi.org/10.1016/j.jksus.2014.11.002>.
- [17]. Arruebo, M.; Fernández-Pacheco, R.; Ibarra, M.R. (2007). Santamaría, J. Magnetic nanoparticles for drug delivery. *Nano Today*, 2, 22–32. [https://doi.org/10.1016/S1748-0132\(07\)70084-1](https://doi.org/10.1016/S1748-0132(07)70084-1).
- [18]. Nemi Malhotra, Jiann-Shing Lee, Rhenz Alfred D. Liman, Johnsy Margotte S. Ruallo, Oliver B. Villaflores, Tzong-Rong Ger, and Chung-Der Hsiao (2020). Potential Toxicity of Iron Oxide Magnetic Nanoparticles: A Review., *Molecules* 2020, 25, 3159. <https://doi.org/10.3390/molecules25143159>
- [19]. Khalil, M. I. (2015). Co-precipitation in aqueous solution synthesis of magnetite Nanoparticles using iron (III) salts as precursors. *Arabian Journal of Chemistry*, 8(2), 279-284. doi:10.1016/j.arabjc.2015.02.008.
- [20]. Suriyaprabha R., Samreen Heena Khan, Bhawana Pathak, and Fulekar M. H. (2017). Spherical Surfaced Magnetic (Fe₃O₄) Nanoparticles as Nano Adsorbent Material for Treatment of Industrial Dye Effluents., *Int. J. Nanosci. Nanotechnol.*, Vol. 13, No. 2, June 2017, pp. 169-175.
- [21]. Luck, H. (1974): *Methods of enzymatic analysis*. Academic press. 885- 894.
- [22]. Yen.P. Yew, K. Shameli, M. Miyake, N. Kuwano, N.B.B.A. Khairudin, S.E.B. Mohamad, K.X. Lee, (2016). Green synthesis of magnetite (Fe₃O₄) nanoparticles using seaweed (*Kappaphycus alvarezii*) extract, *Nanoscale research letters*, vol.11, pp. 1-7.
- [23]. Thomas, J.A.; Schnell, F.; Kaveh-Baghbaderani, Y.; Berensmeier, S.; Schwaminger, S.P. (2020). Immunomagnetic separation of microorganisms with iron oxide nanoparticles. *Chemosensors*, 8, 17. <https://doi.org/10.3390/chemosensors8010017>.
- [24]. Poedji Loekitowati Hariani, Muhammad Faizal, Ridwan, Marsi and Dedi Setiabudidaya (2013). Synthesis and Properties of Fe₃O₄ Nanoparticles by Co-precipitation Method to Removal of Procion Dye. *International Journal of Environ. Sci. and Develop.*, 4(3):366-340.
- [25]. Lida H, Takayanagi K, Nakanishi T, Osaka T. (2007): Synthesis of Fe₃O₄ Nanoparticles with Various Sizes and Magnetic Properties by Controlled Hydrolysis. *J of Colloid and Interface Sci.*, 314(1):274-280. <https://doi.org/10.1016/j.jcis.2007.05.047>.
- [26]. Setiadi, E. A., Sebayang, P., Ginting, M., Sari, A. Y., Kurniawan, C., Saragih, S. ., & Simamora, P. (2016). The synthesization of Fe₃O₄ Magnetic Nanoparticles Based on Natural Iron Sand by Coprecipitation Method for the Used of the Adsorption of Cu and Pb Ions. *Journal of Physics: Conference Series*. <https://doi.org/-10.1088/1742-6596/776/1/012020>.



- [27]. Ferni Malega, I Putu Tedy Indrayana, Edi Suharyadi (2018). Synthesis and Characterization of the microstructure and functional group bond of Fe₃O₄ nanoparticles from natural Iron sand In Tobelo North Halmahera., Jurnal Ilmiah Pendidikan Fisika Al-BiRuNi 07 (2) (2018) 13-22 <https://doi.org/10.24042/jipfalbiruni.v7i2.2913>.
- [28]. Shaw B. J, Handy R. D (2011): Physiological effects of nanoparticles on fish: a comparison of nanometals versus metal ions. Environ Int 37(6):1083–1097. <https://doi.org/10.1016/j.envint.2011.03.009>.
- [29]. Arora, S., Rajwade, J. M., & Paknikar, K. M. (2012). Nanotoxicology and in vitro studies: the need of the hour. Toxicology and applied pharmacology, 258(2), 151-165. <https://doi.org/10.1016/j.taap.2011.11.010>.
- [30]. Dissanayake, N. M., Current, K. M., & Obare, S. O. (2015). Mutagenic effects of iron oxide Nanoparticles on biological cells. International journal of molecular sciences, 16 (10), 23482-23516. <https://doi.org/10.3390/ijms161023482>.
- [31]. Wu, S., Sun, A., Zhai, F., Wang, J., Xu, W., Zhang, Q., & Volinsky, A. A. (2011). Fe₃O₄ Magnetic Nanoparticles synthesis from tailings by ultrasonic chemical co-precipitation. Materials Letters, 65(12), 1882-1884. <https://doi.org/10.1016/j.matlet.2011.03.065>.
- [32]. Saddam Saqib, M.F.H. Munis, W. Zaman, F. Ullah, S.N. Shah, A. Ayaz, M. Farooq, S. Bahadur, (2019). Synthesis, characterization and use of iron oxide nano particles for antibacterial activity, Microscopy research and technique, vol.82, pp. 415–420. <https://doi.org/10.1002/jemt.23182>.
- [33]. Rahdar A, Taboada P, Aliahmad M, Hajinezhad MR, Sadeghfah F. Iron oxide nanoparticles: synthesis, physical characterization, and intraperitoneal biochemical studies in *Rattus norvegicus*. J Mol Struct. 2018;1173:240–5. <https://doi.org/10.1016/j.molstruc.2018.06.098>.
- [34]. Pourmadadi M, Rahmani E, Shamsabadipour A, Mahtabian S, Ahmadi M, Rahdar A, Díez-Pascual AM. Role of iron oxide (Fe₂O₃) nanocomposites in advanced biomedical applications: a state-of-the-art review. Nanomaterials. 2022;12(3873):1–29. <https://doi.org/10.3390/nano12213873>.
- [35]. Taimoory SM, Rahdar A, Aliahmad M, Sadeghfah F, Hajinezhad MR, Jahantigh M, Shahbazi P, Trant JF. The synthesis and characterization of a magnetite nanoparticle with potent antibacterial activity and low mammalian toxicity. J Mol Liq. 2018;265:96–104. <https://doi.org/10.1016/j.molliq.2018.05.105>.
- [36]. Vakili-Ghartavol R, Momtazi-Borojeni AA, Vakili-Ghartavol Z, Aiyelabegan HT, Jaafari MR, Rezayat SM, Arbabi Bidgoli S. Toxicity assessment of superparamagnetic iron oxide nanoparticles in different tissues. Artif Cells, Nanomed Biotech. 2020;48(1):443–51. <https://doi.org/10.1080/21691401.2019.1709855>.
- [37]. Kumari, K., Khare, A., & Dange, S. (2014). The applicability of oxidative stress biomarkers in assessing chromium induced toxicity in the fish *Labeo rohita*. BioMed research international, 2014. BioMed Research International, Article ID 782493, 11, <https://doi.org/10.1155/2014/782493>.
- [38]. Zhu, Z., Tian, S., Cai, Z., (2012). Toxicity assessment of Iron Oxide nanoparticles in zebrafish (*Danio rerio*) early life stages. PLoS ONE 7 (9), 46286.
- [39]. Ates, M., Demir, V., Arslan, Z., Kaya, H., Yulmaz, S., & Camas, M. (2016). Chronic exposure of tilapia (*Oreochromis niloticus*) to iron oxide nanoparticles: Effects of particle morphology on accumulation, elimination, hematology and immune responses. Aquatic toxicology, 177, 22-32. <https://doi.org/10.1016/j.aquatox.2016.05.005>
- [40]. Srikanth, K., Trindade, T., Duarte, A.C., Pereira, E. (2017): Cytotoxicity and oxidative stress responses of silica-coated Iron Oxide nanoparticles in CHSE-214 cells. Environ. Sci. Pollut. Res., 24, 2055–2064.
- [41]. Zheng, M.; Lu, J.; Zhao, D. (2018): Effects of starch-coating of magnetite nanoparticles on cellular uptake, toxicity and gene expression profiles in adult zebrafish. Sci. Total Environ., 622–623, 930–941. <https://doi.org/10.1016/j.scitotenv.2017.12.01>

