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# A Review on Extraction, Isolation and Separation technique of Alkaloids from Tridax Procumbens

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Abstract: Tridax procumbens is a common medicinal herb which is best known as a widespread weed and pest plant distributed throughout India. The chemical constituents of the plant showed that its leaves contain various alkaloids, flavonoids, carotenoids, fumaric acid etc. Their extract possesses various pharmacological properties (anti-inflammatory, hepatoprotective, immunomodulatory, antimicrobial or antibacterial, activity antiseptic, anti-cancerous activity, repellency activity, hemostatic activity, antidiabetic, anti-urolithiatic activity, hypotensive, antioxidant effect, bradycardiac effects etc.). A number of researchers are working on its active constituents in various fields to develop it as a natural ayurvedic medicine against various ailments and disorders. This research work has been undertaken to extract any new or known chemical constituent of Tridax Procubens.

Keywords: Tridax procumbens, Extract, anticoagulant, insect repellant

# I. INTRODUCTION

Tridax procumbens is a species of flowering plant belonging to family Asteraceae and is the most potent species among 30 species. It is best known as widespread weed and pest plant. It is native to the tropical Americas but it has been introduced to tropical, subtropical and mild temperate regions worldwide. It is listed as a noxious weed in the United States and has a pest status. Some of the medicinally important species of the genus Tridax are: T. angustifolia, T. serboana, T. bicolor, T. accedens, T. dubia, T. erecta and T. rosea. procumbens, commonly known as coat buttons or tridax daisy, is aspecies of flowering plant in the daisy family. It is best known as a widespread weedand pest plant. It is native to the tropical Americas but it has been introduced to tropical, subtropical, and mild temperate regions worldwide. Traditionally, Tridax procumbens has been in use in India for wound healing, asanticoagulant, antifungal and insect repellent. It is used in diarrhoea and dysentery. Itsleaf extracts were known to treat infectious skin diseases in folk medicines. It is a well-known ayurvedic medicine for liver disorders or hepato-protective nature besides gastritis and heart burn. A study was carried out to verify the claims wherein tribalinhabitants of Udaipur district, Rajasthan were using the plant for treatment of diabetes. It was concluded that the results were comparable to that of reference standard Glibenclamide and the Tridax procumbens flower extract showed antidiabetic

# TRADITIONAL USES

Traditionally, Tridaxprocumbens has been in use in India for wound healing and as ananticoagulant, antifungal, and insect repellent. The juice extracted from the leaves isdirectly applied on wounds. Its leaf extracts were used for infectious skin diseases infolk medicines. It is used in Ayurvedic medicine for liver disorders, hepato protection, gastritis, and heartburn. Tridax procumbens is also used as treatment for boils, blisters, and cuts by local healers in parts of Indi

# **II. EXTRACTION TECHNIQUES**

Extraction is a critical initial step in isolating bioactive compounds from Coriander Several techniques have been utilized, each with its advantages and limitations, depending on the target compounds and the desired yield.

1. Maceration: Maceration is one of the oldest and simplest extraction methods, involving soaking plant materials in solvents at ambient temperature. The process is straightforward and requires minimal equipment, making it accessible. However, maceration has drawbacks, such as long extraction times and the potential for lower yields. For Moringa

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oleifera, maceration is effective in extracting phenolic compounds, though it may require high solvent volumes and longer durations to achieve satisfactory yields.

# **Procedure of Maceration**

- The plant material is finely ground or crushed to increase the surface area for better solvent penetration.
- The material is then submerged in a solvent such as water, ethanol, methanol, or a solvent mixture.
- The solvent is kept in contact with the material for an extended period, typically ranging from a few hours to several days, depending on the type of material, solvent, and target compounds.
- Stirring or occasional shaking may be applied to enhance the extraction process.
- After maceration, the solvent containing the extracted compounds (the filtrate) is separated from the solid plant material by filtration or decantation.
- The solvent is often evaporated to obtain a concentrated extract, which can then be used for further studies or applications.



# **Maceration Process**

**2. Soxhlet Extraction:** This method involves a continuous solvent reflux, where the solvent repeatedly passes through the plant material, enhancing extraction efficiency. Soxhlet extraction is widely used due to its high yield potential for heat-stable compounds. However, it requires long extraction times and large amounts of organic solvents, which may limit its environmental sustainability and cost-effectiveness. Soxhlet extraction has been effectively used to isolate various bioactive compounds from Tridax Procumbens, including essential oils and other phenolic constituents.

# **Process:**

- The round-bottom flask containing the solvent is heated, causing the solvent to evaporate and travel upwards.
- The vapor reaches the condenser, where it cools and condenses, then drips into the extractor chamber containing the plant material.
- The solvent gradually fills the chamber, allowing the target compounds to dissolve into the solvent.
- Once the chamber reaches a certain level, it siphons back down to the flask, carrying the dissolved compounds with it.
- cycle repeats continuously, with fresh solvent contacting the plant material until the extraction process is complete.

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# 3. Ultrasound-Assisted Extraction (UAE)

UAE utilizes ultrasonic waves to disrupt cell walls, enhancing the release of bioactive compounds. The method is particularly advantageous due to its speed, low solvent requirement, and ability to extract heat-sensitive compounds, as it operates at lower temperatures. UAE has shown high efficacy in extracting antioxidants, phenolics, and essential oils from Tridax procumbens, making it a suitable choice for preserving delicate compounds while reducing extraction time and energy costs.

# Procedure of Ultrasound-Assisted Extraction

- Preparation: The plant material is usually ground or cut into smaller pieces to increase surface area.
- Sonication Setup: The sample is placed in an extraction vessel with a suitable solvent, and an ultrasonic probe or bath generates ultrasonic waves.
- **Extraction Process:** Ultrasonic waves are applied for a specified duration, typically ranging from a few minutes to an hour, depending on the plant material, solvent, and target compounds.
- **Filtration and Concentration:** After extraction, the solvent containing the dissolved compounds is filtered to separate the solid residue. The solvent may then be evaporated to yield a concentrated extract.

# 4. Microwave-Assisted Extraction (MAE)

MAE employs microwave energy to heat solvents and plant materials, improving the efficiency of the extraction process. The method is known for its rapid processing times, reduced solvent usage, and high yields of bioactive compounds. MAE has been successfully applied in extractingtridax procumbensbioactive compounds, such as glucosinolates and saponins, due to its ability to disrupt plant cell matrices and release intracellular compounds effectively. One limitation is the potential degradation of heat-sensitive components if the temperature is not carefully controlled.

# **Procedure of Microwave-Assisted Extraction**

- Preparation: The plant material is often dried and ground to increase surface area.
- Microwave Setup: A microwave reactor or microwave-assisted extraction system is used, containing the plant material and solvent in an extraction vessel.
- Extraction Process: The vessel is subjected to microwave irradiation, rapidly heating the solvent and plant matrix for a specified time (usually a few minutes).
- Filtration and Concentration: After extraction, the solvent is filtered to separate the plant residue. The solvent may then be evaporated or further processed to concentrate the extract.

# 5. Supercritical Fluid Extraction (SFE)

SFE uses supercritical fluids, typically carbon dioxide ( $CO_2$ ), to extract non-polar compounds, offering selectivity, rapid extraction times, and minimal solvent residue. SFE is particularly effective for extracting lipophilic compounds such as essential oils and lipids tridax procumbens , and  $CO_2$ 's non-toxic and non-flammable nature makes it an environmentally friendly choice. However, SFE requires specialized equipment and may have limited effectiveness for polar compounds unless co-solvents are used.

# **Procedure of Supercritical Fluid Extraction**

- Preparation: The plant material is dried and ground to increase the extraction surface area.
- Supercritical Setup: The material is placed in an extraction chamber, and CO<sub>2</sub> is pumped under high pressure, becoming supercritical in state.
- Extraction Process: Supercritical CO<sub>2</sub> passes through the plant matrix, dissolving target compounds. The CO<sub>2</sub> and extracted compounds then flow into a separation chamber, where CO<sub>2</sub> is depressurized back to its gaseous state.

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• Collection and Recovery: As CO<sub>2</sub> becomes a gas again, it releases the extracted compounds, which can then be collected. The CO<sub>2</sub> can be recycled for additional extractions, making the process environmentally efficient.

# 6. Hydro distillation

Commonly used for extracting essential oils, hydrodistillation involves passing steam through the plant material to vaporize volatile compounds, which are then condensed and collected. Although efficient for isolating essential oils, hydrodistillation is limited in extracting non-volatile bioactive compounds. For tridaxprocumbens, hydrodistillation has been used to extract essential oils from leaves, though it may not be ideal for other bioactive.

- Preparation: Plant material is often dried and sometimes chopped or ground to improve extraction efficiency.
- Loading: The plant material is loaded into a distillation chamber with either water or directly exposed to steam, depending on the specific method.
- Heating and Extraction: The chamber is heated, releasing essential oils and volatile compounds as steam.
- Condensation and Separation: The vaporized essential oils and water pass through a condenser, turning into a liquid that collects in a separator. Since essential oils are typically immiscible with water, they form a separate layer that can be easily isolated.

# **Isolation and Separation Techniques**

Once the compounds are extracted, further isolation and separation are required to purify specific bioactive compounds. Techniques used for Moringa oleifera include chromatography, electrophoresis, and other advanced methods, each with varying degrees of specificity and resolution.

**1. Chromatography:** Chromatography techniques, including thin-layer chromatography (TLC), high-performance liquid chromatography (HPLC), and gas chromatography (GC), are widely used for separating and purifying bioactive compounds. HPLC is particularly effective in isolating Moringa compounds due to its high resolution and versatility in separating different types of compounds based on polarity and molecular weight. TLC is often used for initial screening and rapid qualitative analysis, while GC is suitable for volatile compounds such as essential oils. Chromatographic methods have been widely applied in isolating alkaloids ,flavonoidsand phenolic from tridax procumbensextracts.

**2. Electrophoresis:** Electrophoretic techniques, such as capillary electrophoresis (CE), offer high-resolution separation based on the charge-to-size ratio of molecules. Capillary electrophoresis is suitable for analyzing tridax procumbens smaller bioactive compounds and offers rapid analysis with minimal sample and solvent requirements. However, it may be less effective for larger, non-ionic compounds, limiting its application for certain tridax procumbens constituents.

**3. Liquid-Liquid Extraction (LLE):** LLE separates compounds based on their solubility in different solvents. It is a simple yet effective technique for fractionating coriander extracts into polar and non-polar components. LLE has been used as a preliminary separation step before further purification, particularly for isolating hydrophilic and lipophilic compounds in Coriander extracts.

**4. Preparative HPLC:** This technique is an advanced form of HPLC used to isolate large quantities of purified compounds for further study or application. Preparative HPLC has proven effective in isolating Coriander Citratus bioactive constituents with high purity, though it requires sophisticated equipment and expertise.

# **III. CONCLUSION**

Isolation and purification of alkaloids is the difficulty and key of new Chinese medicine research and development, some of the traditional separation and purification technology problem is low yield, high cost of purification. This article describes a method for the extraction of alkaloids in traditional, as opposed to traditional methods, new extraction methods such as membrane separation technology, molecular distillation technology, high-speed countercurrent chromatography, molecular imprinting technology has unparalleled advantages, in improving the alkaloid preparations quality, increasing productivity and quality, reducing environmental pollution, saving time and energy has a positive role in promoting. How these new technologies can be used into production, many problems need

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to be solved. With the development of technology, alkaloids extraction and purification technology will also bemore indepth research and development, we believe that more efficient, convenient and fast method will continue to emerge.

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#### REFERENCES

- [1]. Jadhav V. D., Talele Swati G.\*, BakliwalAkshada A., Chaudhari G. N. SandipInstitute of Pharmaceutical Sciences, Mahiravani, Nasik, India
- [2]. M. V. Sudharani1\*, A. C. Kullayappa1, C. Dheeraj1, K. Bhaskar Naik1, M.Vandana1, P.Jamalbi1 and V. Sravani1
- [3]. D.S. Mohale\*, A.R. Pokrna, C.V. Sanghani, S.R. Rasekar, A.S. Rathi, A.S. Rathod, G. R. Mehetre, A.V. Chandewar\* P. Wadhwani College of Pharmacy, Yavatmal-445001, Maharashtra, India
- [4]. Rajaram S. Sawant and Ashvin G. Godghate\* Department of Botany &Department of Chemistry\* Dr. Ghali College, Gadhinglaj, 416502, Dist. Kolhapur,(M.S.) India
- [5]. SnehaMundada\*, RuchiShivhare S.G.S.P.S., Institute of Pharmacy, Kaulkhed, Akola, HingnaPhata, 444004, M.S., India
- [6]. Abundant seed production. (13) PROPAGATION: Choudhari MM, Maheshwari J.K. (2009) Ethanobotany in South Asia, Middle East. Journal of ScientificResearch 4: 144-146
- [7]. Manjamalia A, Sardar S. S. R., Guruvayooroppan C, Berlin GVM (2010) Analysisof phytochemical constituents and antimicrobial activity of some medicinalplants. Global Journal of Biotechnology and Biochemistry 5: 120-128.
- [8]. Prabhu V V, Nalini G, Chidambaranathan N, Kisan S S. Evaluation of Anti-Inflammatory and Analgesic Activity of Tridax procumbens Linn AgainstFormalin, Acetic Acid and CFA Induced Pain Models, International Journal ofPharmacy and Pharmaceutical Sciences, 3 (2), 2011, 126-130.
- [9]. ChavanChetan, TamboliAshpak, Patil Priyanka, ChavanAshwini,Pharmacognostical and Pharmacological screening of Tridax procumbens,International research journal of pharmacy, 2011; 2 (7): 154-159.
- [10]. GanjuKuldeep, Pathak AK, Pharmacognostic and Phytochemical Evaluation of Tridax procumbens Linn., Journal of Pharmacognosy and Phytochemistry, 2013;1(5): 42.
- [11]. Wani, Minal; Pande, Snehal; More, Nitin (2010). "Callus induction studies inTridax procumbens L."(http://oaji.net/articles/2014/321394172633.pdf)
- [12]. Nallella, Sreeramulu; Suthari, Sateesh; Ragan, A; Raju, Vatsavaya S(2013)."Ethno-botanico-medicine for common human ailments in Nalgonda andWarangal districts of Telangana, Andhra Pradesh, India"(http://annalsofplantsciences.com/index.php/aps/article/view/56). Annals of PlantSciences. 2 (7): 220– 9.6.
- [13]. C.Ikewuchi Jude, C.Ikewuchi, Catherine and M. IgbohNgozi. Chemical ProfileofTridaxprocumbens Linn. Pakistan Journal of Nutrition, 2009, 8 (5), 548-550.
- [14]. R. K. Verma and M. M. Gupta. Lipid constituents of Tridax procumbens, Phytochemistry, 1988, 27 (2), 459-163.
- [15]. Muhammad Shaiq Ali, Muhammad Jahangir, Syed ShazadulHussan, IqbalChoudhary. Inhibition of aglucosidase by oleanolic acid and its Syntheticderivatives. Phytochemistry, 2002, 60, 295–299.
- [16]. FunmilayaAdelowo and Oluwoleoladeji, Department of pune andappliedchemistry, laolokeAkintold, University of Technology, P. M. B. 4000,



