

# Unveiling the Power Within: A Review of Postbiotics as Dynamic Agents with Antimicrobial Capabilities

Mahaur Kamlesh Ramdas<sup>1</sup> and Dr. Patel Bhavik Ambulal<sup>2</sup>

Research Scholar, Department of Pharmaceutics<sup>1</sup>

Research Guide, Department of Pharmaceutics<sup>2</sup>

Sunrise University, Alwar, Rajasthan, India

**Abstract:** *The concept of postbiotics, representing the diverse array of bioactive molecules produced by beneficial microorganisms during fermentation or metabolic processes, has emerged as a promising avenue in the field of antimicrobial research. These dynamic biological molecules, derived from the byproducts of probiotics or the microbial communities inhabiting the human body, exhibit considerable potential in modulating and enhancing the host's immune response. This abstract delves into the multifaceted role of postbiotics as potent agents with antimicrobial activity, elucidating their mechanisms of action and therapeutic implications. By exploring the intricate interplay between postbiotics and host microbiota, this review aims to shed light on the evolving landscape of postbiotic research and its implications for developing innovative strategies in combating microbial infections and promoting overall health.*

**Keywords:** Dynamic biological molecules, Antimicrobial activity, Bioactive compounds

## I. INTRODUCTION

Postbiotics were just discovered, making probiotics hard to get. Postbiotic supplements should include unique postbiotics, such as short-chain fatty acids [1], which may replace some nutrients in diets. Human gut microbiotas are diverse, and postbiotics may maintain their balance [2].

Probiotics are beneficial microorganisms that may benefit health. Probiotics in food or the host body may create postbiotics, which cause most of these health effects. However, probiotic function's molecular pathways are complex and unknown [3]. Despite probiotics' beneficial role in the gut, absorption, infection risk, and antibiotic resistance gene transfer remain concerns [4]. Optimizing probiotic therapy involves increasing probiotic survivability in fermented food and as they transit through the human gastrointestinal tract to the colon, their main action site, to 10<sup>7</sup> cfu/g or ml. Probiotics support prebiotics during fermentation, which may be a byproduct of postbiotics, and may help keep the stomach healthy by avoiding unwanted microbes [5]. Studies suggest postbiotics may prevent type 1 diabetes in pre-diabetics [6]. A parenthetically unbalanced microbiota may cause pre-diabetes or metabolic resistance [7].

Muramyl Dipeptide (MDP), a postbiotic, has been proven to diminish affront resistance even after weight reduction or gut flora changes during obesity [8]. Schertzer concluded that microbial imbalance and postbiotic synthesis disturbances contribute to diabetes [9]. Changing the number and diversity of indigenous bacteria in the gastrointestinal tract may affect type 2 diabetes [10]. The gut bacteria's blood sugar-lowering mechanism is well recognized [11]. Karim et al. (2017) evaluated how inulin RG14 and postbiotics affect development, cecal smaller-scale biota, unstable greasy volatile fatty acids, and optimum cytokine explanation in broilers [12].

The study found no difference between Interleukin 8/Chemokine (C-X-C motif) value and slim down. In aviculture, inulin (prebiotic) and postbiotics (produced from probiotics) may replace antimicrobial development boosters [13]. Konstantinos et al. [14] concluded that postbiotics may be safer than live bacteria for colonic health. Tsilengiri et al. suggested using postbiotics in therapy and anticipation with gut-relevant illnesses like provocative intestine illness [16]. Studies have shown that probiotics that produce post-antibiotics like exopolysaccharides can improve gastrointestinal health and prevent colon and rectal cancer [15].

An effective 2017 assessment reviewed randomized, controlled human studies with any clinical outcome that included a butchered probiotic as a mediation [17]. Audit questions numbered forty. These 40 proposals differed in organism, target population, and end purpose (preventing or curing a cluster of illnesses). However, creators say thoughts were probably not aroused to notice a change. Dead probiotics outperformed live ones in two therapeutic tests [18]. One avoidance approach said life was better than murder. The audit also looked for dead organism side effects. As usual, most ponders unsuccessfully gathered or described competing circumstances to avoid a conclusion [19].

### **After biotics**

Postbiotics are beneficial bioactive complexes that are created in a fermentation environment and may be used to enhance consumer health. In the fermentation environment, a wider variety of postbiotics are created the more prebiotic carbohydrates that probiotics have access to [20]. According to some research, probiotics may not always be able to support and sustain intestinal health on their own, but the postbiotics they create may be able to do so successfully [21]. Probiotics create many postbiotics, such as lactospin, indole, lipopolysaccharide, exopolysaccharide, teichoic acid, and muramyl dipeptide. Acetic acid, butyric acid, and propionic acid are examples of short-chain fatty acids that are among the most important and well studied postbiotics [22]. These fats are essential to the intestines' physiological and digestive processes. Maintaining gut health is mostly dependent on taking part in certain metabolic processes as well as food digestion and absorption [23]. As probiotic postbiotics, short-chain carbohydrates improve health by promoting the growth or activity of one or more gut microbiota and are unaffected by human stomach-related proteins [23–24]. These substances, which come from *Lactobacillus* species, operate as a stand-in for live probiotic cells in food and the body when there are insufficient probiotics [18]. A probiotic bacteria may produce postbiotic, a metabolic byproduct that affects the host's organic capacity [25].

Stout people with pre-diabetes may benefit from the aid of bacterial by-products, sometimes known as postbiotics, in decreasing their blood sugar levels [26]. Researchers hypothesize that postbiotics might help overweight people with pre-diabetes avoid developing type 2 diabetes [26–27]. Postbiotics are now thought to be helpful prebiotics that, in addition to their own positive effects, nourish and boost the efficacy of probiotics as a rich supply of carbohydrates [28]. The pH is lowered and the environment for the development and activity of pathogenic bacteria are restricted by the synthesis of postbiotics, particularly short-chain fatty acids [29].

Postbiotics are essential for promoting *Lactobacillus* and *Bifidobacterium* bacterial growth and activity [30]. It was previously believed that the existence and activity of bacteria increased blood sugar and inflammatory responses. Nevertheless, it is now shown that probiotic bacteria and postbiotic substances lower blood sugar and enhance insulin action in obese individuals [31]. In a study where scientists genetically modified fat mice, they discovered that postbiotics boost the effects of insulin [32–35].

### **Antimicrobial Activity of Postbiotics**

Postbiotics address the bacterial risks of live cell probiotics by arguing that bacterial life is not essential for human health [36]. Despite little study, postbiotics may have antimicrobial properties [37]. Research has demonstrated that postbiotic compounds have antibacterial (pathogenic and spoiler) effects that prevent food spoilage and infectious diseases. These compounds prevent intestinal problems including diarrhea and irritable bowel syndrome by inhibiting gut bacteria [38]. Postbiotics improve human health by changing the body's microbial ecology, limiting antimicrobial growth, replacing invading gut bacteria, and changing environmental pH [39]. By maintaining intestinal microbiota balance and starting fermentation and post-antibiotic production, intestinal infections and other gastrointestinal tract problems are considerably reduced [40, 41]. Postbiotics connect to bacterial receptors to limit the development of toxic pathogens like *E. coli* and *Clostridium* and remove them from the intestinal lumen [42, 43].

Recent research on postbiotics' positive benefits and relevant ingredients has shown that they may prevent surviving microbes or premature babies from replacing and causing illness [44, 45].

### **Microbial Strains Used as Postbiotic Sources**

This may be important to avoid the creation of safe antibacterial microbial strains that might damage humans [46]. *Lactobacillus* strains connected to people must be employed as postbiotics in animals, whereas *Bifidobacterium* strains

from humans were used [47]. Bacillus strains have been used to make postbiotics in most countries, notably Europe, for decades [48]. The main end products of postbiotics are exopolysaccharides and short-chain volatile fatty acids synthesized in the lab. Most postbiotics used in breeding are synthetic and given to birds, pigs, and ruminants [49, 50, 51]. These compounds may also be found in pig, ruminant, and chick feces [53, 54, 55]. Bacillus subtilis When Mind 588 is separated from ocean water, E. coli may thrive [56].

Giang et al. recovered lactic acid bacteria (LAB) from sound stuffing pig interiors. Postbiotics such Pediococcus pentosaceus, L. acidophilus, L. plantarum, and Enterococcus faecium helped weaned pigs develop [56–57]. Silages from humid and hot environments have isolated Pediococcus pentosaceus, P. lolii, L. pentosus, L. plantarum, L. buchneri, L. rafi, and L. rhamnosus [58, 59]. Some study suggests that L. murinus, L. salivarius, and L. johnsonii from young feces calves may become bacteriocins [60].

Some investigations have identified postbiotic strains different from newly found aquatic and marine organisms [61]. L. salivarius from bottlenose porpoises may prevent human and marine Salmonella enteritidis strains from spreading [62]. Pig small intestines yielded particular Lactobacillus strains, mostly L. salivarius [63]. These organisms showed potential postbiotic traits such pH 3 tolerance, auto-aggregation, and E. coli K88 manifestation [64].

Leuconostoc mesenteroides was found in Nile tilapia and snakehead fish intestines [65, 53]. Common octopuses Leuconostoc cremoris and Weissella cibaria of Atlantic salmon fish provided postbiotics [66].

Sarkono et al. found that L. paracasei from the normal condition resists bile and acidic environments and kills pathogenic bacteria including E. Coli, Bacillus cereus, and Staphylococcus aureus [67].

Leuconostoc mesenteroides strains have been obtained from the intestines employing new water angles such Nile tilapia [69–71] and snakehead fish [68].

**Table Some bacterial activities species of postbiotics.**

Competition with pathogen	References
Lactobacillus plantarum I-UL4	[61]
Lactobacillus rhamnosus	[66]
Lactobacillus paracasei	[68]
Faecalibacterium prausnitzii	[68]
Lactobacillus brevis	[70]
Lactobacillus pentosus	[70]
Lactobacillus gasser	[71]

**Postbiotics as Antimicrobial Agents in Food Products**

LAB are confirmed safe, active, and useful food components with a long history of usage [72]. They may also employ their metabolic byproducts, such as lactic acid and bacteriocin, as conventional viewpoint and bactericidal agents to prevent food rotting [73].

LAB prevents urogenital infections [74], controls inflammatory bowel diseases [75], modulates the immune system [76], regulates serum cholesterol [77], and inhibits certain cancers [78]. Bio-preserving cell-free supernatant from L. plantarum YML 007 increased unshelled soybean shelf life by two months [79]. Exopolysaccharide from L. rhamnosus boosted cheddar cheese production by 8.2% when coupled with L. lactis [68]. Bifidin from Bifidobacterium lactis Bb-12 increased minced beef shelf life to three months at -18°C by decreasing E. Coli O157:H7 by 100% [80].

Many practical issues hinder good bacterial cell effects [81]. However, undefined terms like "postbiotic" or "paraprobiotic" imply that dead microorganisms, microbial divisions, or cell lysates may indicate physiological preferences for bioactivity [82]. Numerous investigations on LAB postbiotics have demonstrated that each genus and species produces a different soluble postbiotic. Many of these bacteria develop mucoid colonies and films in the culture medium [83]. L. plantarum, the main LAB strain, produces PM with unique postbiotic effects [84]. More is known about LAB's anticancer activities, but less about PM from L. plantarum's cytotoxic and anti-proliferative actions. Research has been done on the cytotoxicity of PM produced by six L. plantarum strains on normal and malignant cells [85].

**Antimicrobial mechanisms of postbiotics**

It has been examined and shown that postbiotic metabolites have a good potential to substitute in-feed antimicrobials in animal nutrition [86]. Postbiotics aim to mimic the viable restorative effects, predicting the risk of preserving living bacteria in prematurely born infants with immature intestinal barriers or impaired immune systems [87]. Many microbes in the gastrointestinal system have the ability to catabolize indigestible carbohydrates to create significant quantities of volatile short-chain fatty acids and butyrate [89].

**Here are some of the health-promoting effects of postbiotics:**

Due to poor carbohydrate fermentation, water is retained in the gastrointestinal tract, causing constipation. The colon's capacity to absorb fibrous foods boosts stool output and bacteria [90]. In an aged study, inulin, an indigestible carbohydrate/prebiotic, reduced constipation and increased feces [91]. Carbohydrate postbiotics like exopolysaccharides operate similarly.

Postbiotics lower animal blood lipids, according to research. But further study is needed to confirm this effect on humans [92]. Postbiotics affect triglyceride activity and fatty acid synthesis, although human studies have not been done [93].

Postbiotics change and strengthen gut flora, boosting the body's and gastrointestinal tract's immune systems and lowering the risk of inflammatory illnesses, especially intestinal inflammation [94].

Human and animal studies have indicated that postbiotics improve iron, calcium, magnesium, and zinc absorption [95]. Human experiments have revealed that digestible oligosaccharides improve calcium absorption during adolescence and menstruation. Increase intestinal lumen calcium, potassium, and magnesium ions to control and inhibit cellular transformation [96].

Effect on cancer: As probiotics in the gut expand, the body creates more postbiotics such as butyrate and oligosaccharides, which prevent cancer and cell deformation [97]. Postbiotics may increase Lactobacillus and Bifidobacteria activity, causing them to bind and destroy carcinogens [98]. Postbiotic highlights' appropriate inhibitory effect is crucial to their mechanical growth. Bacitracin synthesis under controlled aging has occurred for decades [99].

**Future Perspectives of Postbiotics**

Postbiotic antibacterial properties and microbial resistance are discussed in this article. Due to antibiotics' strengthening effects, co-administration of antibiotics and postbiotics may help combat germ resistance. This idea needs further laboratory and clinical animal and human study [100, 101]. The most notable distinction between postbiotics and probiotics is that probiotics must be active [102]. Live bacteria or dead and damaged cell components secrete postbiotics. Postbiotics are particularly interesting because of how much simpler fabrication, packaging, packing, transportation, and caring are without the organism. Since probiotics' active nature may cause infection concerns, inactive and dead organisms are a safer alternative. Post-biotics have been demonstrated to improve health in several research. Postbiotics may be microencapsulated to improve effectiveness and survivability like probiotics [103, 104].

**II. CONCLUSION**

These days, probiotic effects ascertained by means of microbial metabolites regarded as bioactive postbiotic metabolites are receiving more and more attention. Using living microorganisms, postbiotics are defined as dissolvable agents (items or metabolic by-products) that are released after bacterial lysis, such as proteins, teichoic acids, peptides, peptidoglycans, cell surface proteins, polysaccharides, and natural acids. The self-evident chemical features, security dosage items, long shelf life, and the content of various signaling particles that will have anti- (inflammatory, obesogenic, immunomodulatory, hypertensive, proliferative, oxidant) hypocholesterolemia applications have drawn interest in these postbiotics. Even though the specifics are still unclear, these targeted benefits of postbiotics may contribute to the advancement of wellbeing by boosting the execution of particular physiological wants.

**REFERENCES**

- [1]. Usha, G.; Ravi, D.; Parthasarathy, R. Synthesis of Bacteriocin by Synbiotic Effect and Its Antibacterial Activity against Selected Respiratory tract Pathogens. *Int J Adv Res* 2013, 1, 296-303, <http://www.journalijar.com/article/644/>.
- [2]. Rossoni, R.D.; de Barros, P.P.; do Carmo Mendonça, I.; Medina, R.P.; Silva, D.H.S.; Fuchs, B.B.; Junqueira, J.C.; Mylonakis, E. The postbiotic activity of *Lactobacillus paracasei* 28.4 against *Candida auris*. *Front Cell Infect Microbiol* 2020, 10, 397, <https://doi.org/10.3389/fcimb.2020.00397>.
- [3]. Goudarzi, L.; Kasra Kermanshahi, R. The Effect of Prebiotics on Production of Antimicrobial Compounds from *Lactobacillus* spp. Against *Proteus mirabilis* (ATCC 7002 and PTCC 1076). *Iran Food Sci Technol Res J* 2015, 11, 41-47. [www.sid.ir/en/journal/ViewPaper.aspx?id=535997](http://www.sid.ir/en/journal/ViewPaper.aspx?id=535997).
- [4]. Bindels, L.B.; Delzenne, N.M.; Cani, P.D.; Walter, J. Towards a more comprehensive concept for prebiotics. *Nat. Rev. Gastroenterol. Hepatol.* 2015, 12, 303-310, <https://doi.org/10.1038/nrgastro.2015.47>.
- [5]. Nat. Rev. Gastroenterol. Hepatol. 2015, 12, 303-310, <https://doi.org/10.1038/nrgastro.2015.47>.
- [6]. Izuddin, W.I.; Humam, A.M.; Loh, T.C.; Foo, H.L.; Samsudin, A.A. Dietary Postbiotic *Lactobacillus plantarum* Improves Serum and Ruminant Antioxidant Activity and Upregulates Hepatic Antioxidant Enzymes and Ruminant Barrier Function in Post-Weaning Lambs. *Antioxidants* 2020, 9, 250, <https://doi.org/10.3390/antiox9030250>.
- [7]. Kumar, M.; Nagpal, R.; Verma, V.; Kumar, A.; Kaur, N.; Hemalatha, R.; Gautam, S.K.; Singh, B. Probiotic metabolites as epigenetic targets in the prevention of colon cancer. *Nutr. Rev.* 2013, 71, 23-34, <https://doi.org/10.1111/j.1753-4887.2012.00542.x>.
- [8]. Kareem, K.Y.; Hooi Ling, F.; Teck Chwen, L.; Foong, O.M.; Asmara, S.A. Inhibitory activity of postbiotic produced by strains of *Lactobacillus plantarum* using reconstituted media supplemented with inulin. *Gut Pathog* 2014, 6, 23, <https://doi.org/10.1186/1757-4749-6-23>.
- [9]. Gao, J.; Li, Y.; Wan, Y.; Hu, T.; Liu, L.; Yang, S.; Gong, Z.; Zeng, Q.; Wei, Y.; Yang, W.; Zeng, Z.; He, X.; Huang, S.-H.; Cao, H. A Novel Postbiotic From *Lactobacillus rhamnosus* GG With a Beneficial Effect on Intestinal Barrier Function. *Front. Microbiol.* 2019, 10, 477, <https://doi.org/10.3389/fmicb.2019.00477>.
- [10]. Howarth, G.; Wang, S.H. Role of endogenous microbiota, probiotics and their biological products in human health. *Nutrients* 2013, 5, 58–81, <https://doi.org/10.3390/nu5010058>.
- [11]. Cremon, C.; Barbaro, M.R.; Ventura, M.; Barbara, G. Pre- and probiotic overview. *Cur Opin Pharmacol* 2018, 43, 87-92, <https://doi.org/10.1016/j.coph.2018.08.010>.
- [12]. 2018, 43, 87-92, <https://doi.org/10.1016/j.coph.2018.08.010>.
- [13]. Kareem, K.Y.; Loh, T.C.; Foo, H.L.; Asmara, S.A.; Akit, H. Influence of postbiotic RG14 and inulin combination on cecal microbiota, organic acid concentration, and cytokine expression in broiler chickens. *Poult Sci* 2017, 96, 966–975, <https://doi.org/10.3382/ps/pew362>.
- [14]. Konstantinov, S.; Kuipers, E.; Peppelenbosch, M. Functional genomic analyses of the gut microbiota for CRC screening. *Nat Rev Gastroenterol Hepatol* 2013, 10, 741–745, <https://doi.org/10.1038/nrgastro.2013.178>.
- [15]. Dadi, T.H.; Vahjen, W.; Zentek, J.; Melzig, M.F.; Granica, S.; Piwowski, J.P. *Lythrum salicaria*, L. Herb and gut microbiota of healthy post-weaning piglets. Focus on prebiotic properties and formation of postbiotic metabolites in ex vivo cultures. *J Ethnopharmacol* 2020, 261, 113073, <https://doi.org/10.1016/j.jep.2020.113073>.
- [16]. Sugiharto, S. Role of nutraceuticals in gut health and growth performance of poultry. *J Saudi Soc Agric Sci* 2016, 15, 99-111, <https://doi.org/10.1016/j.jssas.2014.06.001>.
- [17]. 2016, 15, 99-111, <https://doi.org/10.1016/j.jssas.2014.06.001>.
- [18]. Zorzela, L.; Ardestani, S.K.; McFarland, L.V.; Vohra, S. Is there a role for modified probiotics as beneficial microbes: a systematic review of the literature. *Benef Microbes* 2017, 8, 739-754, <https://doi.org/10.3920/BM2017.0032>.
- [19]. Arena, M.P.; Silvain, A.; Normanno, G.; Grieco, F.; Drider, D.; Spano, G.; Fiocco, D. Use of *Lactobacillus plantarum* Strains as a Bio-Control Strategy against Food-Borne Pathogenic Microorganisms. *Front. Microbiol.* 2016, 7, 464, <https://doi.org/10.3389/fmicb.2016.00464>.
- [20]. Kuitunen, M.; Kukkonen, A.K.; Savilahti, E. Impact of Maternal Allergy and Use of Probiotics during Pregnancy on Breast Milk Cytokines and Food Antibodies and Development of Allergy in Children until 5 Years. *Int Arch Allergy Immunol* 2012, 159, 162-170, <https://doi.org/10.1159/000336157>.

- [21]. Wu, X.; Teame, T.; Hao, Q.; Ding, Q.; Liu, H.; Ran, C.; Yang, Y.; Zhang, Y.; Zhou, Z.; Duan, M.; Zhang, Z. Use of a paraprobiotic and postbiotic feed supplement (HWFT<sup>TM</sup>) improves the growth performance, composition and function of gut microbiota in hybrid sturgeon (*Acipenser baerii* x *Acipenser schrenckii*). *Fish Shellfish Immunol.* 2020, 104, 36-45, <https://doi.org/10.1016/j.fsi.2020.05.054>.
- [22]. Kasra Kermanshahi, R. Goodarzi, L.; The Effect of Prebiotics on Production of Antimicrobial Compounds from *Lactobacillus* spp. Against *Proteus mirabilis* (ATCC 7002 and PTCC 1076). *Iran J Food Sci Technol Res* 2015, 11, 41-47, <https://doi.org/10.22067/ijfstrj.v11i1.45434>.
- [23]. Pérez-Sánchez, T.; Mora-Sánchez, B.; Vargas, A.; Balcázar, J.L. Changes in intestinal microbiota and disease resistance following dietary postbiotic supplementation in rainbow trout (*Oncorhynchus mykiss*). *Microb pathog* 2020, 142, 104060, <https://doi.org/10.1016/j.micpath.2020.104060>.
- [24]. Ford, A.C.; Harris, L.A.; Lacy, B.E.; Quigley, E.M.; Moayyedi, P. Systematic review with meta-analysis: the efficacy of prebiotics, probiotics, synbiotics and antibiotics in irritable bowel syndrome. *Aliment pharmacol ther* 2018, 48, 1044-1060, <https://doi.org/10.1111/apt.15001>.
- [25]. Hayes, S.R.; Vargas, A.J. Probiotics for the prevention of pediatric antibiotic-associated diarrhea. *Explore* 2016, 12, 463-466, <https://doi.org/10.1016/j.explore.2016.08.015>.
- [26]. Holanda, D.M.; Yiannikouris, A.; Kim, S.W. Investigation of the efficacy of a postbiotic yeast cell wall-based blend on newly-weaned pigs under a dietary challenge of multiple mycotoxins with emphasis on deoxynivalenol. *Toxins* 2020, 12, 504, <https://doi.org/10.3390/toxins12080504>.
- [27]. Al-Sheraji, S.H.; Ismail, A.; Manap, M.Y.; Mustafa, S.; Yusof, R.M.; Hassan, F.A. Prebiotics as functional foods: A review. *J funct foods* 2013, 5, 1542-1553, <https://doi.org/10.1016/j.jff.2013.08.009>.
- [28]. Ooi, M.F.; Foo, H.L.; Loh, T.C.; Mohamad, R.; Rahim, R.A.; Ariff, A. A refined medium to enhance the antimicrobial activity of postbiotic produced by *Lactiplantibacillus plantarum* RS5. *Sci. Rep.* 2021, 11, 7617, <https://doi.org/10.1038/s41598-021-87081-6>.
- [29]. Inglin, R.C.; Stevens, M.J.A.; Meile, L.; Lacroix, C.; Meile, L. High-throughput screening assays for antibacterial and antifungal activities of *Lactobacillus* species. *J. Microbiol. Methods* 2015, 114, 26-29, <https://doi.org/10.1016/j.mimet.2015.04.011>.
- [30]. Patel, R.M.; Denning, P.W. Therapeutic use of prebiotics, probiotics, and postbiotics to prevent necrotizing enterocolitis: what is the current evidence? *Clin Perinatol* 2013, 40, 11-25, <https://doi.org/10.1016/j.clp.2012.12.002>.
- [31]. Żółkiewicz, J.; Marzec, A.; Ruszczyński, M.; Feleszko, W. Postbiotics-A Step Beyond Pre-and Probiotics. *Nutrients* 2020, 12, 2189, <https://doi.org/10.3390/nu12082189>.
- [32]. Izuddin, W.I.; Loh, T.C.; Samsudin, A.A.; Foo, H.L. In vitro study of postbiotics from *Lactobacillus plantarum* RG14 on rumen fermentation and microbial population. *Revista Brasileira de Zootecnia* 2018, 47, <https://doi.org/10.1590/rbz4720170255>.
- [33]. Loh, T.C.; Choe, D.W.; Foo, H.L.; Sazili, A.Q.; Bejo, M.H. Effects of feeding different postbiotic metabolite combinations produced by *Lactobacillus plantarum* strains on egg quality and production performance, faecal parameters and plasma cholesterol in laying hens. *BMC Vet. Res.* 2014, 10, 149, <https://doi.org/10.1186/1746-6148-10-149>.
- [34]. Martens, E.C.; Kelly, A.G.; Tauzin, A.S.; Brumer, H. The devil lies in the details: how variations in polysaccharide fine-structure impact the physiology and evolution of gut microbes. *J Molecul Biol* 2014, 426, 3851-3865, <https://doi.org/10.1016/j.jmb.2014.06.022>.
- [35]. Yordshahi, A.S.; Moradi, M.; Tajik, H.; Molaie, R. Design and preparation of antimicrobial meat wrapping nanopaper with bacterial cellulose and postbiotics of lactic acid bacteria. *Int J Food Microbiol* 2020, 321, 108561, <https://doi.org/10.1016/j.ijfoodmicro.2020.108561>.
- [36]. Holscher, H.D.; Caporaso, J.G.; Hooda S.; Jennifer M.; Brulc, G.C.; Fahey, J.R.; Swanson, K.S. Fiber supplementation influences phylogenetic structure and functional capacity of the human intestinal microbiome: follow-up of a randomized controlled trial. *Am J clin nutr* 2015, 101, 55-64, <https://doi.org/10.3945/ajcn.114.092064>.

- [39]. Oliver, L.; Heather, R.; Mary B.G.; Yimin, C. Health care provider's knowledge, perceptions, and use of probiotics and prebiotics. *Top Clin Nutr* 2014, 29, 139-149, <https://doi.org/10.1097/01.TIN.0000445898.98017.eb>.
- [40]. Pandey, K.R.; Naik, S.R.; Vakil, B.V. Probiotics, prebiotics and synbiotics- a review. *J Food Sci Technol* 2015, 52, 7577-7587, <https://doi.org/10.1007/s13197-015-1921-1>.
- [41]. Ridwan, B. U.; Koning, C. J. M.; Besselink, M. G. H.; Timmerman, H. M.; Brouwer, E. C.; Verhoef, J.; Akkermans, L. M. A. Antimicrobial activity of a multispecies probiotic (Ecologic 641) against pathogens isolated from infected pancreatic necrosis. *Letters Appl Microbiol* 2008, 46, 61-67, <https://doi.org/10.1111/j.1472-765X.2007.02260.x>.
- [42]. Moradi, M.; Kousheh, S. A.; Almasi, H.; Alizadeh, A.; Guimarães, J. T.; Yilmaz, N.; Lotfi, A. Postbiotics produced by lactic acid bacteria: The next frontier in food safety. *Com Rev Food Sci Food Saf* 2020, 19, 3390-3415, <https://doi.org/10.1111/1541-4337.12613>.
- [43]. Chen, C.C.; Lai, C.C.; Huang, H.L.; Huang, W.Y.; Toh, H.S.; Weng, T.C.; Chuang, Y.C.; Lu, Y.C.; Tang, H.J. Antimicrobial Activity of Lactobacillus Species Against Carbapenem-Resistant Enterobacteriaceae. *Front Microb* 2019, 10, 789, <https://doi.org/10.3389/fmicb.2019.00789>.
- [44]. Tang, H.-J.; Hsieh, C.-F.; Chang, P.-C.; Chen, J.-J.; Lin, Y.-H.; Lai, C.-C.; Chao, C.-M.; Chuang, Y.-C. Clinical Significance of Community- and Healthcare-Acquired Carbapenem-Resistant Enterobacteriaceae Isolates. *PLoS One* 2016, 11, e0151897, <https://doi.org/10.1371/journal.pone.0151897>.
- [45]. Majeed, M.; Majeed, S.; Nagabhushanam, K.; Mundkur, L.; Rajalakshmi, H.R.; Shah, K.; Beede, K. Novel Topical Application of a Postbiotic, LactoSporin, in Mild to Moderate Acne: A Randomized, Comparative Clinical Study to Evaluate its Efficacy, Tolerability and Safety. *Cosmetics* 2020, 7, 70, <https://doi.org/10.3390/cosmetics7030070>.
- [46]. Shah, N.; Patel, A.; Ambalam, P.; Holst, O.; Ljungh, A.; Prajapati, J. Determination of an antimicrobial activity of Weissella confusa, Lactobacillus fermentum, and Lactobacillus plantarum against clinical pathogenic strains of Escherichia coli and Staphylococcus aureus in co-culture. *Ann. Microbiol.* 2016, 66, 1137-1143, <https://doi.org/10.1007/s13213-016-1201-y>.
- [47]. Brown, D.F.J.; Wootton, M.; Howe, R.A. Antimicrobial susceptibility testing breakpoints and methods from BSAC to EUCAST. *J. Antimicrob. Chemother.* 2016, 71, 3-5, <https://doi.org/10.1093/jac/dkv287>.
- [48]. Tokatli, M.; Gulgor, G.; Bagder Elmaci, S.; Arslankoz İsleyen, N.; Özçelik, F. In vitro properties of potential probiotic indigenous lactic acid bacteria originating from traditional pickles. *Biomed Res Int* 2015, 315819, <https://doi.org/10.1155/2015/315819>.
- [49]. Tang, H.-J.; Lai, C.-C.; Chen, C.-C.; Zhang, C.-C.; Weng, T.-C.; Chiu, Y.-H.; Toh, H.-S.; Chiang, S.-R.; Yu, W.-L.; Ko, W.-C.; Chuang, Y.-C. Colistin-sparing regimens against Klebsiella pneumoniae carbapenemase-producing K. pneumoniae isolates: Combination of tigecycline or doxycycline and gentamicin or amikacin. *J. Microbiol. Immunol. Infect.* 2019, 52, 273-281, <https://doi.org/10.1016/j.jmii.2016.03.003>.
- [50]. Humam, A.M.; Loh, T.C.; Foo, H.L.; Samsudin, A.A.; Mustapha, N.M.; Zulkifli, I.; Izuddin, W.I. Effects of feeding different postbiotics produced by Lactobacillus plantarum on growth performance, carcass yield, intestinal morphology, gut microbiota composition, immune status, and growth gene expression in broilers under heat stress. *Animals* 2019, 9, 644, <https://doi.org/10.3390/ani9090644>.
- [51]. Loh, T. C.; Thu, T. V.; Foo, H. L.; Bejo, M. H. Effects of different levels of metabolite combination produced by Lactobacillus plantarum on growth performance, diarrhoea, gut environment and digestibility of postweaning piglets. *J Appl Animal Res* 2013, 41, 200-207, <https://doi.org/10.1080/09712119.2012.741046>.
- [52]. Jean, S.S.; Coombs, G.; Ling, T.; Balaji, V.; Rodrigues, C.; Mikamo, H. Epidemiology and antimicrobial susceptibility profiles of pathogens causing urinary tract infections in the Asia-Pacific region: results from the study for monitoring antimicrobial resistance trends (SMART), 2010-2013. *Int J Antimicrob Agents* 2016, 47, 328-334, <https://doi.org/10.1016/j.ijantimicag.2016.01.008>.
- [53]. Ajuwon, K. M. Toward a better understanding of mechanisms of probiotics and prebiotics action in poultry species. *J Appl Poult Res* 2016, 25, 277-283, <https://doi.org/10.3382/japr/pfv074>.
- [54]. Ebrahimi, M.; Sadeghi, A.; Rahimi, D.; Purabdollah, H.; Shahryari, S. Postbiotic and Anti-aflatoxigenic

- Capabilities of *Lactobacillus kunkeei* as the Potential Probiotic LAB Isolated from the Natural Honey. *Probiotics and Antimicrobial Proteins* 2021, 13, 343-355, <https://doi.org/10.1007/s12602-020-09697-w>.
- [56]. Rodriguez-Bano, J.; Gutierrez-Gutierrez, B.; Machuca, I.; Pascual, A. Treatment of infections caused by extended-spectrum-beta-lactamase-, ampC-, and carbapenemase-producing Enterobacteriaceae. *Clin Microb Rev* 2018, 31, e00079-17, <https://doi.org/10.1128/CMR.00079-17>.
- [57]. Păcularu-Burada, B.; Georgescu, L. A.; Vasile, M. A.; Rocha, J. M.; Bahrim, G. E. Selection of wild lactic acid bacteria strains as promoters of postbiotics in gluten-free sourdoughs. *Microorganisms* 2020, 8, 643. <https://doi.org/10.3390/microorganisms8050643>.
- [58]. Kumar, M.; Dhaka, P.; Vijay, D.; Vergis, J.; Mohan, V.; Kumar, A.; Kurkure, N.V.; Barbudhe, S.B.; Malik, S.V.S.; Rawool, D.B. Antimicrobial effects of *Lactobacillus plantarum* and *Lactobacillus acidophilus* against multidrug-resistant enteroaggregative *Escherichia coli*. *Int. J. Antimicrob. Agents* 2016, 48, 265-270, <https://doi.org/10.1016/j.ijantimicag.2016.05.014>.
- [59]. Teame, T.; Wang, A.; Xie, M.; Zhang, Z.; Yang, Y.; Ding, Q.; Gao, C.; Olsen, R.E.; Ran, C.; Zhou, Z. Paraprobiotics and Postbiotics of Probiotic Lactobacilli, Their Positive Effects on the Host and Action Mechanisms: A Review. *Front Nutr* 2020, 7, 570344, <https://doi.org/10.3389/fnut.2020.570344>.
- [60]. Kang, M. S.; Lim, H. S.; Oh, J. S.; Lim, Y. J.; Wuertz-Kozak, K.; Harro, J. M.; Achermann, Y. Antimicrobial activity of *Lactobacillus salivarius* and *Lactobacillus fermentum* against *Staphylococcus aureus*. *Pathog Dis* 2017, 75, <https://doi.org/10.1093/femspd/ftx009>.
- [61]. Mookiah, S.; Sieo, C. C.; Ramasamy, K.; Abdullah, N.; Ho, Y. W. Effects of dietary prebiotics, probiotic and synbiotics on performance, caecal bacterial populations and caecal fermentation concentrations of broiler chickens. *J Sci of Food Agric* 2014, 94, 341-348, <https://doi.org/10.1002/jsfa.6365>.
- [62]. Georgieva, R.; Yocheva, L.; Tserovska, L.; Zhelezova, G.; Stefanova, N.; Atanasova, A.; Karaivanova, E. Antimicrobial activity and antibiotic susceptibility of *Lactobacillus* and *Bifidobacterium* spp. intended for use as starter and probiotic cultures. *Biotechnol Biotechnol Equip* 2015, 29, 84-91, <https://doi.org/10.1080/13102818.2014.987450>.
- [63]. Cortés-Martín, A.; Selma, M. V.; Tomás-Barberán, F. A.; González-Sarrias, A.; Espín, J. C. Where to look into the puzzle of polyphenols and health? The postbiotics and gut microbiota associated with human metabolotypes. *Molecul Nutr Food Res* 2020, 64, 1900952, <https://doi.org/10.1002/mnfr.201900952>.
- [64]. Grimoud, J.; Durand, H.; Courtin, C.; Monsan, P.; Ouarné, F.; Theodorou, V.; Roques, C. In vitro screening of probiotic lactic acid bacteria and prebiotic glucooligosaccharides to select effective synbiotics. *Anaerobe* 2010, 16, 493-500, <https://doi.org/10.1016/j.anaerobe.2010.07.005>.
- [65]. Stein, DR.; Allen, DT.; Perry, EB.; Bruner, JC.; Gates, KW.; Rehberger, T.; Mertz, K.; Jones, D.; Spicer, LJ. Effects of feeding propionic bacteria to dairy cows on milk yield, milk components, and reproduction. *J Dairy Sci* 2008, 9, 111-125, [https://doi.org/10.3168/jds.S0022-0302\(06\)72074-4](https://doi.org/10.3168/jds.S0022-0302(06)72074-4).
- [66]. Rad, A.H.; Maleki, L.A.; Kafil, H.S.; Zavoshti, H.F.; Abbasi, A. Postbiotics as novel health-promoting ingredients in functional foods. *Health Promot Perspect* 2020, 10, 3-4, <https://doi.org/10.15171/hpp.2020.02>.
- [67]. Prieto, M. L.; O'Sullivan, L.; Tan, S. P.; McLoughlin, P.; Hughes, H.; Gutierrez, M.; Gardiner, G. E. In vitro assessment of marine *Bacillus* for use as livestock probiotics. *Mar Drugs* 2014, 12, 2422-2445, <https://doi.org/10.3390/md12052422>.
- [68]. Chuah, L.-O.; Foo, H.L.; Loh, T.C.; Mohammed Alitheen, N.B.; Yeap, S.K.; Abdul Mutalib, N.E.; Abdul Rahim, R.; Yusoff, K. Postbiotic metabolites produced by *Lactobacillus plantarum* strains exert selective cytotoxicity effects on cancer cells. *BMC Complement. Altern. Med.* 2019, 19, 114, <https://doi.org/10.1186/s12906-019-2528-2>.
- [69]. Malagón-Rojas, J.N.; Mantziari, A.; Salminen, S.; Szajewska, H. Postbiotics for Preventing and Treating Common Infectious Diseases in Children: A Systematic Review. *Nutrients* 2020, 12, <https://doi.org/10.3390/nu12020389>.
- [70]. Devi, S.M.; Halami, P.M. Genetic Variation of pln Loci Among Probiotic *Lactobacillus plantarum* Group Strains with Antioxidant and Cholesterol-Lowering Ability. *Probiotics Antimicrob* 2019, 11, 11-22, <https://doi.org/10.1007/s12602-017-9336-0>.



- [71]. Koohestani, M.; Moradi, M.; Tajik, H.; Badali, A. Effects of cell-free supernatant of *Lactobacillus acidophilus* LA5 and *Lactobacillus casei* 431 against planktonic form and biofilm of *Staphylococcus aureus*. *Vet Res forum* 2018, 9, 301–306, <https://doi.org/10.30466/vrf.2018.33086>.
- [72]. Carlsson, A. H.; Yakymenko, O.; Olivier, I.; Håkansson, F.; Postma, E.; Keita, Å. V.; Söderholm, J. D. Faecalibacterium prausnitzii supernatant improves intestinal barrier function in mice DSS colitis. *Scan J gastroenterol* 2013, 48, 1136-1144, <https://doi.org/10.3109/00365521.2013.828773>.
- [73]. Rushdy, A.A.; Gomaa, E.Z. Antimicrobial compounds produced by probiotic *Lactobacillus brevis* isolated from dairy products. *Ann Microb* 2013, 63, 81–90, <https://doi.org/10.1007/s13213-012-0447-2>.
- [74]. Muhialdin, B.J.; Hassan, Z.; Sadon, S.K. Antifungal Activity of *Lactobacillus fermentum* Te007, *Pediococcus pentosaceus* Te010, *Lactobacillus pentosus* G004, and *L. paracasi* D5 on Selected Foods. *J Food Sci* 2011, 76, M493-M499, <https://doi.org/10.1111/j.1750-3841.2011.02292.x>.
- [75]. Tejero-Sariñena, S.; Barlow, J.; Costabile, A.; Gibson, G. R.; Rowland, I. In vitro evaluation of the antimicrobial activity of a range of probiotics against pathogens: evidence for the effects of organic acids. *Anaerobe* 2012, 18, 530-538, <https://doi.org/10.1016/j.anaerobe.2012.08.004>.
- [76]. Maldonado, N.C.; de Ruiz, C.S.; Otero, M.C.; Sesma, F.; Nader-Macías, M. E. Lactic acid bacteria isolated from young calves—characterization and potential as probiotics. *Res Vet sci* 2012, 92, 342-349, <https://doi.org/10.1016/j.rvsc.2011.03.017>.
- [77]. Ang, C.Y.; Sano, M.; Dan, S.; Leelakriangsak, M.; Lal, T.M. Postbiotics applications as infectious disease control agent in aquaculture. *Biocontrol Sci* 2020, 25, 1-7, <https://doi.org/10.4265/bio.25.1>.
- [78]. Khorshidian, N.; Yousefi Asli, M.; Hosseini, H.; Shadnoush, M.; Mortazavian, A. M. Potential anticarcinogenic effects of lactic acid bacteria and probiotics in detoxification of process-induced food toxicants. *Iran J Cancer Prevent* 2016, 1-13, <http://eprints.semums.ac.ir/id/eprint/908>.
- [79]. Giang, H.H.; Viet, T.Q.; Ogle, B.; Lindberg, J.E. Effects of different probiotic complexes of lactic acid bacteria on growth performance and gut environment of weaned piglets. *Livestock Sci* 2010, 133, 182-184, <https://doi.org/10.1016/j.livsci.2010.06.059>.
- [80]. Doi, K.; Nishizaki, Y.; Kimura, H.; Kitahara, M.; Fujino, Y.; Ohmomo, S.; Ogata, S. Identification of thermo tolerant lactic acid bacteria isolated from silage prepared in the hot and humid climate of Southwestern Japan. *Springerplus* 2013, 2, 1-12, <https://doi.org/10.1186/2193-1801-2-485>.
- [81]. Boricha, A. A.; Shekh, S. L.; Pithva, S. P.; Ambalam, P. S.; Vyas, B. R. M. In vitro evaluation of probiotic properties of *Lactobacillus* species of food and human origin. *LWT-Food Sci Technol* 2019, 106, 201-208, <https://doi.org/10.1016/j.lwt.2019.02.021>.
- [82]. Monteagudo-Mera, A.; Rodríguez-Aparicio, L.; Rúa, J.; Martínez-Blanco, H.; Navasa, N.; García-Armesto, M.R.; Ferrero, M.Á. In vitro evaluation of physiological probiotic properties of different lactic acid bacteria strains of dairy and human origin. *J Funct Foods* 2012, 4, 531-541, <https://doi.org/10.1016/j.jff.2012.02.014>.
- [83]. Vrzáčková, N.; Ruml, T.; Zelenka, J. Postbiotics, Metabolic Signaling, and Cancer. *Molecules* 2021, 26, 1528, <https://doi.org/10.3390/molecules26061528>.
- [84]. Devi, S.M.; Archer, A.C.; Halami, P.M. Screening, Characterization and In Vitro Evaluation of Probiotic Properties Among Lactic Acid Bacteria Through Comparative Analysis. *Probiotics Antimicrob* 2015, 7, 181-192, <https://doi.org/10.1007/s12602-015-9195-5>.
- [85]. Allameh, S.K.; Daud, H.; Yusoff, F.M.; Saad, C.R.; Ideris, A. Isolation, identification and characterization of *Leuconostoc mesenteroides* as a new probiotic from intestine of snakehead fish (*Channa striatus*). *African J Biotechnol* 2012, 11, 3810-3816, <https://doi.org/10.5897/AJB11.1871>.
- [86]. Zapata, A. A.; Lara-Flores, M. Antimicrobial activities of lactic acid bacteria strains isolated from Nile Tilapia intestine (*Oreochromis niloticus*). *J Biol Life Sci* 2013, 4, 164-171, <https://doi.org/10.5296/jbls.v4i1.2408>.
- [87]. Muñoz-Atienza, E.; Araújo, C.; Magadán, S.; Hernández, P.E.; Herranz, C.; Santos, Y.; Cintas, L.M. In vitro and in vivo evaluation of lactic acid bacteria of aquatic origin as probiotics for turbot (*Scophthalmus maximus* L) farming. *Fish Shellfish Immunol* 2014, 41, 570-580, <https://doi.org/10.1016/j.fsi.2014.10.007>.

- [88]. Moreno Munoz, J. A.; Chenoll, E.; Casinos, B.; Bataller, E.; Ramón, D.; Genovés, S.; Rivero, M. Novel probiotic *Bifidobacterium longum* subsp. *infantis* CECT 7210 strain active against rotavirus infections. *Appl Environ Microbiol* 2011, 77, 8775-8783, <https://doi.org/10.1128/AEM.05548-11>.
- [89]. Sarkono, S.; Faturrahman F.; Sofyan Y. Isolation and identification of lactic acid bacteria from abalone (*Haliotis asinina*) as a potential candidate of probiotic. *Nus Biosci* 2010, 2, 2087-3940, <https://doi.org/10.13057/nusbiosci/n020106>.
- [90]. Karovičová, J.; Kohajdová, Z. Lactic acid fermentation of various vegetable juices. *Acta Aliment* 2005, 34, 237-246, <https://doi.org/10.1556/aalim.34.2005.3.5>.
- [91]. Jama, Y.H.; Varadaraj, M. Antibacterial effect of plantaricin LP84 on foodborne pathogenic bacteria occurring as contaminants during idli batter fermentation. *World J Microb Biotechnol* 1999, 15, 27-32, <https://doi.org/10.1023/A:1008887201516>.
- [92]. Mantziari, A.; Salminen, S.; Szajewska, H.; Malagón-Rojas, J. N. Postbiotics against pathogens commonly involved in pediatric infectious diseases. *Microorganisms* 2020, 8, 1510, <https://doi.org/10.3390/microorganisms8101510>.
- [93]. Lammers, K.M.; Vergopoulos, A.; Babel, N.; Gionchetti, P.; Rizzello, F.; Morselli, C.; Caramelli, E.; Fiorentino, M.; d'Errico, A.; Volk, H.-D.; Campieri, M. Probiotic Therapy in the Prevention of Pouchitis Onset: Decreased Interleukin-1 $\beta$ , Interleukin-8, and Interferon- $\gamma$  Gene Expression. *Inflamm. Bowel Dis.* 2005, 11, 447-454, <https://doi.org/10.1097/01.mpa.0000160302.40931.7b>.
- [94]. Kato-Mori, Y.; Orihashi, T.; Kanai, Y.; Sato, M.; Sera, K.; Hagiwara, K. Fermentation metabolites from *Lactobacillus gasseri* and *Propionibacterium freudenreichii* exert bacteriocidal effects in mice. *J Med Food* 2010, 13, 1460-1467, <https://doi.org/10.1089/jmf.2010.1137>.
- [95]. Fong, W.; Li, Q.; Yu, J. Gut microbiota modulation: a novel strategy for prevention and treatment of colorectal cancer. *Oncogene* 2020, 39, 4925-4943, <https://doi.org/10.1038/s41388-020-1341-1>.
- [96]. Ratajczak, C.; Duez, C.; Grangette, C.; Pochard, P.; Tonnel, A. B.; Pestel, J. Impact of lactic acid bacteria on dendritic cells from allergic patients in an experimental model of intestinal epithelium. *J Biomed Biotechnol* 2007, 2007, 71921, <https://doi.org/10.1155/2007/71921>.
- [97]. Kießling, G.; Schneider, J.; Jahreis, G. Long-term consumption of fermented dairy products over 6 months increases HDL cholesterol. *Eur. J. Clin. Nutr.* 2002, 56, 843-849, <https://doi.org/10.1038/sj.ejcn.1601399>.
- [98]. Ishikawa, H.; Akedo, I.; Otani, T.; Suzuki, T.; Nakamura, T.; Takeyama, I.; Kakizoe, T. Randomized trial of dietary fiber and *Lactobacillus casei* administration for prevention of colorectal tumors. *Int J Cancer* 2005, 116, 762-767, <https://doi.org/10.1002/ijc.21115>.
- [99]. Ahmad Rather, I.; Seo, B.; Rejish Kumar, V.J.; Choi, U.H.; Choi, K.H.; Lim, J.H.; Park, Y.H. Isolation and characterization of a proteinaceous antifungal compound from *Lactobacillus plantarum* YML 007 and its application as a food preservative. *Letters Appl Microbiol* 2013, 57, 69-76, <https://doi.org/10.1111/lam.12077>.
- [100]. Torino, M.I.; Font de Valdez, G.; Mozzi, F. Biopolymers from lactic acid bacteria. Novel applications in foods and beverages. *Front Microbiol* 2015, 6, 834, <https://doi.org/10.3389/fmicb.2015.00834>.
- [101]. Bosch, M.; Nart, J.; Audivert, S.; Bonachera, M.A.; Alemany, A.S.; Fuentes, M.C.; Cuné, J. Isolation and characterization of probiotic strains for improving oral health. *Arch Oral Biol* 2012, 57, 539-549, <https://doi.org/10.1016/j.archoralbio.2011.10.006>.
- [102]. Aguilar-Toalá, J.E.; Garcia-Varela, R.; Garcia, H.S.; Mata-Haro, V.; González-Córdova, A.F.; Vallejo-Cordoba, B.; Hernández-Mendoza, A. Postbiotics: An evolving term within the functional foods field. *Trends Food Sci Technol* 2018, 75, 105-114, <https://doi.org/10.1016/j.tifs.2018.03.009>.
- [103]. Wegh, C. A.; Geerlings, S.Y.; Knol, J.; Roeselers, G.; Belzer, C. Postbiotics and their potential applications in early life nutrition and beyond. *Int J Mol Sci* 2019, 20, 4673, <https://doi.org/10.3390/ijms20194673>.
- [104]. Reynés, B.; Palou, M.; Rodríguez, A.M.; Palou, A. Regulation of adaptive thermogenesis and browning by prebiotics and postbiotics. *Front physiol* 2019, 9, 1908, <https://doi.org/10.3389/fphys.2018.01908>.