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## Review of Strategies for the Industrial Production of α-amylase by *Bacillus subtilis*

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**Abstract:** Bacillus subtilis can secrete industrially important proteins such as proteases and  $\alpha$ -amylases and used on industrial scale.  $\alpha$ -Amylase enzyme has market demands due to its applications in food, bakery, detergent industries, starch liquefaction, pre-digestion of the animal feed to enhance its quality, sizing of the fibres in textiles. The safety issues associated with the use of this bacteria for industrial applications are studied and it has been observed that the products obtained from it are having GRAS status of US Food and Drug Administration. Bacillus subtilis is considered the most widely experimental organism to conduct the genetic modification studies due to its properties which make it a suitable host for biosynthesis of the products. The genomic structure of Bacillus subtilis can be modified with the help of high quality genomic sequences. The genetic strategies for such modifications include the use of mutagenic treatments, screening of better expression systems, use of better promoters and high secretion level peptides. Another aspect of these strategies to enhance the enzyme yield includes the application of different fermentation methods and use of different substrates. Present review article summarizes some of such strategies applied for obtaining higher yields of  $\alpha$ -amylase enzyme using Bacillus subtilis.

Keywords: Bacillus Subtilis,  $\alpha$ -amylase, Fermentation, Genetic Modification, Screening, Enzyme Production.

## REFERENCES

- [1]. Westers, L., Westers, H., & Quax, W. J. (2004). Bacillus subtilis as cell factory for pharmaceutical proteins: a biotechnological approach to optimize the host organism. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*, 1694(1-3), 299-310.
- [2]. Guan, C., Cui, W., Cheng, J., Zhou, L., Liu, Z., & Zhou, Z. (2016). Development of an efficient autoinducible expression system by promoter engineering in Bacillus subtilis. *Microbial Cell Factories*, 15(1), 1-12.
- [3]. Jaouadi, N. Z., Jaouadi, B., Aghajari, N., & Bejar, S. (2012). The overexpression of the SAPB of Bacillus pumilus CBS and mutated sapB-L31I/T33S/N99Y alkaline proteases in Bacillus subtilis DB430: new attractive properties for the mutant enzyme. *Bioresource technology*, *105*, 142-151.
- [4]. Mountain, A. (1989). Gene expression systems for Bacillus subtilis. In *Bacillus* (pp. 73-114). Springer, Boston, MA.
- [5]. Mulimani VH, Ramalingam Patil GN. Amylase production by solid state fermentation: a new practical approach to biotechnology courses. Biochem Edu 2000; 28:161/3.
- **[6].** Shene, C., Andrews, B. A., & Asenjo, J. A. (1999). Fedbatch fermentations of Bacillus subtilis ToC46 (pPFF1) for the synthesis of a recombinant  $\beta$ -1, 3-glucanase: experimental study and modelling. *Enzyme and microbial technology*, 24(5-6), 247-254.
- [7]. Dawes, I. W., & Thornley, J. H. M. (1970). Sporulation in Bacillus subtilis. Theoretical and experimental studies in continuous culture systems. *Microbiology*, *62*(1), 49-66.
- [8]. Fordyce, A. P., & Rawlings, J. B. (1996). Segregated fermentation model for growth and differentiation of Bacillus licheniformis. *AIChE journal*, 42(11), 3241-3252.

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- [9]. Tiwari, Sp and Srivastava, R and Singh, Cs and Shukla, Kartikeya and Singh, Rk and Singh, Pushpendra and Singh, Ravindra and Singh, Nl and Sharma, Rajesh,(2015), pages (1886-1901), Amylases: an overview with special reference to alpha amylase, volume4, Journal of Global Biosciences
- [10]. Ramachandran, S., Patel, A. K., Nampoothiri, K. M., Chandran, S., Szakacs, G., Soccol, C. R., & Pandey, A. (2004). Alpha amylase from a fungal culture grown on oil cakes and its properties. *Brazilian archives of biology and technology*, 47, 309-317.
- [11]. Goyal, N., Gupta, J. K., & Soni, S. K. (2005). A novel raw starch digesting thermostable α-amylase from Bacillus sp. I-3 and its use in the direct hydrolysis of raw potato starch. *Enzyme and Microbial Technology*, 37(7), 723-734.
- [12]. Sivaramakrishnan, S., Gangadharan, D., Nampoothiri, K. M., Soccol, C. R., & Pandey, A. (2006). a-Amylases from microbial sources-an overview on recent developments. *Food Technol Biotechnol*, 44(2), 173-184.
- **[13].** Andersson, E., Johansson, A. C., & Hahn-Hägerdal, B. (1985). α-Amylase production in aqueous two-phase systems with Bacillus subtilis. *Enzyme and microbial technology*, *7*(7), 333-338.
- [14]. Mosbach, K., Birnbaum, S., Hardy, K., Davies, J., & Bülow, L. (1983). Formation of proinsulin by immobilized Bacillus subtilis. *Nature*, 302(5908), 543-545.
- [15]. Pandey, A., Nigam, P., Soccol, C. R., Soccol, V. T., Singh, D., & Mohan, R. (2000). Advances in microbial amylases. *Biotechnology and applied biochemistry*, 31(2), 135-152.
- [16]. Haddaoui, E; Chambert, R; Petit-Glatron, M.F; Lindy, O; ; Sarvas, M Bacillus subtilis kamylase: The rate limiting step of secretion is growth phase-independent, FEMS Microbiology. Lett. 173 (1999) 127–131.
- [17]. Tanyildizi, M.S.; Ozer, D; Elibol, M., Optimization of α-amylase production by Bacillus sp. using response surface methodology, Process Biochemistry. 40 (2005) 2291–2296
- [18]. Baysal, Z., Uyar, F., & Aytekin, C. (2003). Solid state fermentation for production of  $\alpha$ -amylase by a thermotolerant Bacillus subtilis from hot-spring water. *Process Biochemistry*, *38*(12), 1665-1668.
- [19]. Huang, H., Ridgway, D., Gu, T., & Moo-Young, M. (2003). A segregated model for heterologous amylase production by Bacillus subtilis. *Enzyme and Microbial Technology*, *32*(3-4), 407-413.
- [20]. Konsoula, Z., & Liakopoulou-Kyriakides, M. (2006). Thermostable  $\alpha$ -amylase production by Bacillus subtilis entrapped in calcium alginate gel capsules. *Enzyme and Microbial Technology*, 39(4), 690-696.
- **[21].** Krishna, C., & Chandrasekaran, M. (1996). Banana waste as substrate for α-amylase production by Bacillus subtilis (CBTK 106) under solid-state fermentation. *Applied Microbiology and Biotechnology*, *46*(2), 106-111.
- [22]. Ma, Y., Shen, W., Chen, X., Liu, L., Zhou, Z., Xu, F., & Yang, H. (2016). Significantly enhancing recombinant alkaline amylase production in Bacillus subtilis by integration of a novel mutagenesis-screening strategy with systems-level fermentation optimization. *Journal of biological engineering*, *10*(1), 1-11.
- [23]. Ploss, T. N., Reilman, E., Monteferrante, C. G., Denham, E. L., Piersma, S., Lingner, A., & van Dijl, J. M. (2016). Homogeneity and heterogeneity in amylase production by Bacillus subtilis under different growth conditions. *Microbial cell factories*, 15(1), 1-16.
- **[24].** Rajagopalan, G., & Krishnan, C. (2008). α-Amylase production from catabolite derepressed Bacillus subtilis KCC103 utilizing sugarcane bagasse hydrolysate. *Bioresource technology*, *99*(8), 3044-3050.